

Drivers of Functional Diversity in the Hyporheic Zone of a Large River

Marie-José Dole-Olivier, Michel Creuzé Des Châtelliers, Diana M.P. Galassi, Michel Lafont, Florian Mermillod-Blondin, Frédéric Paran, Didier Graillot, Shishir Gaur, Pierre Marmonier

► To cite this version:

Marie-José Dole-Olivier, Michel Creuzé Des Châtelliers, Diana M.P. Galassi, Michel Lafont, Florian Mermillod-Blondin, et al.. Drivers of Functional Diversity in the Hyporheic Zone of a Large River. Science of the Total Environment, 2022, 843, pp.156985. 10.1016/j.scitotenv.2022.156985. emse-03695444

HAL Id: emse-03695444 https://hal-emse.ccsd.cnrs.fr/emse-03695444v1

Submitted on 21 Nov 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1	
2	
3	
4 5	Drivers of functional diversity in the hyporheic zone of a large river
6	
7	
8	Marie-José Dole-Olivier ^a , Michel Creuzé des Châtelliers ^a , Diana M. P. Galassi ^b , Michel Lafont ^a , Florian
9 10	Mermillod-Blondin", Frederic Paran", Didier Graillot", Shishir Gaur" and Pierre Marmonier" (\Join)
10	
12	a - Univ. Lyon, University Claude Bernard Lyon 1, CNRS, ENTPE, UMR5023 LEHNA, 43 Boulevard du 11
13	Novembre 1918, F-69622, Villeurbanne, France
14	
15	b - University of L'Aquila, Department of Life, Health and Environmental Sciences, Via Vetoio, Coppito,
16	67100 L'Aquila, Italy.
17	
18 10	c – École Nationale des Mines de Saint Étienne, CNRS, UMR5600 PEG, F-42023 Saint Étienne, France
20	d – Department of Civil Engineering, Indian Institute of Technology (BHU), 221005 Varanasi, India.
21	
22	(🖂) Corresponding author:
23	e-mail: <u>pierre.marmonier@univ-lyon1.fr</u>
24	
25	
26	
27	Acknowledgements
28	The work was funded by Electricité de France (convention CNRS-109651) in the framework of the ZABR
29	(LTSER Rhone River Basin). We thank the Ecole Universitaire de Recherche H2O'Lyon, Laurent Cadilhac
30	and Stéphane Stroffek of the Rhone River Water Agency (Agence de l'Eau Rhône Méditerranée et
31	Corse), Céline Thicoïpé of the SR3A (Syndicat de la Rivière d'Ain Aval et de ses Affluents), Hervé Piegay
32	(UMR 5600, EVS), Sylvain Doledec (UMR 5023, LEHNA) and Anne Clémens of the ZABR for discussion
33	and information. Lara Konecny-Dupré (UMR 5023, LEHNA) help us in the molecular analyses of

Clitellata and Niphargides and Bjoern Wissel and Zoraida Quinones-Rivera for comments and English

- editing.

38 Highlights: (3 to 5 point with 85 characters max)

- 39 -Hydrogeology, current and past fluvial forms control large-scale distribution of the hyporheos
- 40 -Hydrology controls hyporheos composition at the station scale
- 41 -Visible characteristics of surface sediment had poor influence
- 42 -Upwelling zones are hotspot of groundwater biodiversity
- 43 -Downwelling zones harboured high abundances of temporary hyporheos

45 Abstract

46 The effects of regional (hydrogeology and geomorphology) and local (sediment and hydrology) 47 characteristics on hyporheic assemblages were studied along a 40-km reach of a large gravel-bed river. Hyporheic water and fauna were sampled at the upstream and downstream positions of 15 large gravel 48 49 bars. The resulting 30 stations varied in their sediment grain size, stability and direction of river-aquifer 50 exchanges. The study concludes that at the 40-km (sector) scale, the longitudinal distribution of 51 hyporheic fauna was controlled by 1) the hydrogeology of the valley (i.e. gaining vs loosing sectors) 52 that modifies abundance and taxonomic richness of stygobites 2) current channel morphometry of the 53 river (i.e. shape and location of meanders), and 3) historical changes (i.e. river incision) which modify 54 abundance and richness of assemblages. At the local scale, we found that surface grain size and stability of the sediment evaluated by visual observation were poor predictors of hyporheos 55 56 composition. In contrast, the local hydrology (i.e. downwellings, upwellings, low vertical exchanges) 57 explained a large part of the abundance, taxonomic richness and composition of the hyporheic 58 assemblages. Stations with low vertical exchanges were found poorly colonized, while the upwelling 59 zones were rich in stygobites and downwelling areas harbor abundant and species-rich temporary 60 hyporheos. It was also observed that functional diversity was controlled by the same parameters, with high relative abundances of stygobites in upwelling zones and POM feeders in downwelling zones. The 61 62 heterogeneity of hydrological patterns, with alternation of upwellings and downwellings may 63 represent the optimal spatial structure for hyporheic biodiversity conservation and resilience in rivers. 64

- 65
- 66

67 Key words: Hyporheos, stygobites, hydrogeology, geomorphology, river incision, dams

- 68 (Six max)
- 69

- 70 **1. Introduction**
- 71

72 Porous sediment deposits in rivers constitute the hyporheic habitat (Orghidan, 1959), where river-73 aquifer exchanges occur together with a large number of ecological processes important for stream 74 metabolism (Findlay, 1995; Boulton et al., 1998). Hyporheic processes contribute to the recycling of 75 nutrients (e.g., nitrification / denitrification, Triska et al., 1993; Jones et al., 1995, Storey et al., 2004) 76 and of organic matter (e.g. biodegradation of leaf litter, Cornut et al., 2010). Additionally, the hyporheic 77 habitat contributes to the biodiversity of the river by combining colonization of both obligate groundwater fauna (i.e. stygobites) and benthic organisms (Dole-Olivier and Marmonier 1992; Dole-78 79 Olivier et al., 1997; Brunke and Gonzer, 1999). Hyporheic organisms can that modify the structure of 80 the sediment (through bioturbation, Mermillod-Blondin and Rosenberg, 2006; Shrivastava et al., 81 2021a, 2021b), the dynamics of organic matter (through subsurface biodegradation, Crenshaw et al., 82 2002; Marmonier et al., 2010) and the concentration of dissolved nutrients (Mermillod-Blondin et al., 83 2000; Nogaro and Burgin, 2014). Thus, their biodiversity is of crucial importance for river integrity and 84 functioning.

85

The hydrological, geomorphological, physicochemical, and/or biological processes control major 86 87 ecological features of streams (Petts & Amoros, 1996). The local hydrology, with the occurrence of 88 water exchanges between the river and the adjacent groundwater is the major driver for the control 89 of hyporheic processes and biodiversity (Dole-Olivier and Marmonier 1992; Valett et al., 1993; Brunke 90 and Gonzer, 1999). Many studies have demonstrated the influence of river-aquifer exchanges on the 91 physicochemical characteristics, nutrient recycling and invertebrate biodiversity of streams (e.g., Olsen 92 and Townsend, 2003; Navel et al., 2011; Mathers et al., 2017). The intensity and direction of these 93 exchanges are crucial; in downwelling zones, the river water infiltrates into the riverbed sediments 94 carrying fresh organic matter and dissolved oxygen, while in upwelling zones, the flow of groundwater 95 towards the surface stabilizes stream temperature and water level (Claret et al., 1998; Franken et al., 96 2001; Peralta-Maraver et al., 2018). Additionally, the direction and magnitude of hydrological 97 exchanges through the hyporheic zone also control hyporheic biodiversity (Dole-Olivier & Marmonier, 98 1992; Lin et al., 2020).

99

These river-aquifer interactions are generally controlled by the geology of the valley and the geomorphology of the streambed at different scales (Williams, 1993; Poole, 2002, 2010; Capderrey et al., 2013). The geology of the valley controls the characteristics of the aquifer (e.g. volume, general slope) and the relative elevation of the piezometric level compared to the river level (Graillot et al., 2014; Marmonier et al., 2019). Similarly, the geomorphology of the streambed controls the water 105 exchanges between the river and the hyporheic zone (Buffington and Tonina, 2009): at a regional scale, 106 the shape of the channel influences the direction of water exchanges (Cardenas et al., 2004). For 107 example, large gravel bars induce infiltration of surface water at their heads and exfiltration of 108 groundwater at their tails (Dole-Olivier and Marmonier, 1992). Furthermore, the geomorphology 109 controls the local characteristics of sediments like their grain sizes and their stability (e.g. Singer, 2008; 110 Snelder et al., 2011). For example, the occurrence of fine sediment patches may modify the nature and intensity of most hyporheic microbial processes, due to reductions in vertical water exchanges, 111 112 availability of dissolved oxygen and organic matter (Lefebvre et al., 2005; Nelson et al., 2019). Likewise, 113 sediment grain size may control hyporheic assemblages, for both taxonomic richness and species 114 composition (Descloux et al., 2013; O'Sullivan et al., 2019) and sediment mobility maintain hyporheic 115 permeability and vertical migration of the hyporheic fauna (Dole-Olivier et al., 1997). The effects of 116 these large-scale characteristics (i.e. hydrogeology and geomorphology) are not limited to hyporheic 117 processes, but also control the global functioning of the river and the dynamics of most organisms that 118 live inside and in close relation to the river, supporting the importance to consider the river in its valley 119 (Hynes, 1975) as a global hydrosystem (Petts & Amoros, 1996).

120

121 Hyporheic assemblages (hyporheos) are characterized by a variety of taxa with very different ecological 122 requirements. Some species use the hyporheic zone for only part of their life cycle (i.e. temporary 123 hyporheos), while other species can spend their entire life cycle within the sediment (i.e. permanent 124 hyporheos). In addition, taxa such as stygobites are obligated to live in groundwater habitats (Williams, 125 1984; Marmonier et al., 1993). The relative composition of these three groups of species may 126 determine zones of high diversity (hyporheic hotspots, Marmonier et al., 2020; Hutchins et al., 2020). 127 Furthermore, the hyporheic fauna is very diverse in food preferences ranging from fine sediment 128 feeders, to particulate organic matter-feeders (detritivores) and predators (Williams et al., 2010). 129 While taxonomic composition of river assemblages may be strongly influenced by biogeography, 130 evolutionary processes and dispersal (Eme et al., 2015), the functional diversity based on ecological 131 groups (Usseglio-Polatera et al., 2000) can help to elucidate the dynamics of hyporheic assemblages.

132

Functional diversity relates to the range and value of those species and organismal traits that influence how an ecosystem functions (Lawton, 1994; Tilman et al., 1997). It may also be affected by large-scale gradients in habitat characteristics (Poff, 1997; Brown et al., 2018) or by local disturbances (Schmera et al., 2017; Graco-Rosa et al., 2021). For example, Parker et al. (2018) used a functional index based on fish habitats and foraging traits to evaluate the effect of long-term improvements in water quality in four reaches of the Illinois River. In the same way, Paillex et al. (2013) used the functional diversity of macroinvertebrate assemblages based on 11 biological traits to estimate the effect of the 140 restoration of the connectivity across the Rhône River floodplain. Also, functional diversity of invertebrates may be of interest to evaluate the effects of disturbances in the hyporheic zone 141 142 (Magliozzi et al., 2019; Di Lorenzo et al., 2021). Changes in direction and/or intensity of rivergroundwater exchanges can lead to shifts in hyporheic biodiversity, whether they are natural (e.g. 143 144 floods) or human-induced (e.g. dewatering, groundwater abstraction; Dole-Olivier et al., 1997; Datry, 145 2012). Similarly, changes in the local characteristics of the bottom sediment (e.g. erosion, sediment 146 clogging) can also modify the composition of hyporheic assemblages (Strayer et al., 1997; Marmonier 147 et al., 2010b), but with poor changes in taxonomic richness due to species replacement (Descloux et al., 2013). Functional diversity -that include ecological requirements of species-may be efficient to 148 149 evaluate the effect of changes in river-groundwater exchanges or in sediment characteristics on 150 hyporheic species composition (Marmonier et al., 2012; Descloux et al., 2014; Lewandowski et al., 151 2019).

152

153 The present work aims to determine the combined effects of regional patterns (i.e. hydrogeological 154 and geomorphological characteristics of a 40-km long reach) and local characteristics (i.e. sediment 155 grain size, sediment stability and river-hyporheic water exchanges) on the composition and the 156 functional diversity of hyporheic assemblages. The local hydrology was described using water 157 chemistry, while the sediment characteristics was assessed by image analyses. The functional diversity 158 was characterized using two functional traits based on the affinity of the species for groundwater 159 habitats and their food-type preferences. For this purpose, 30 stations were selected along 15 gravel 160 bars of a large gravel-bed river (the Ain River, France), with varying river-groundwater exchanges, 161 sediment grain size and stability. Specifically, the following three hypotheses were proposed.

First, hydrogeology and geomorphology control hyporheic assemblages at the scale of the 40 km reach (H1, Poole, 2010). More precisely, both river-groundwater exchanges (obtained from
 hydrogeology modelling) and channel morphology (e.g., location of incision zones, meanders)
 explain the longitudinal variations of hyporheic abundance, its taxonomic richness, and the
 variation of ecological groups along the studied reach.

167 Second, surface sediment characteristics (visual estimation of the grain size composition and stability) have an effect on the composition of the hyporheic assemblages at the local scale 168 169 (H2, Descloux et al., 2013). Taxonomic richness and abundances are high in coarse and mobile 170 sediment, which favor the vertical migration of benthic organisms (temporary hyporheos) and 171 organisms that feed on POM infiltrating from the surface. Whereas organisms that feed on fine sediment and algae are more abundant in sandy and stable areas covered by abundant 172 173 biofilms. Finally, the stygobites that live in deep groundwater and are sensible to environmental instability are more abundant in stable shallow sediments. 174

Third, the vertical hydrological exchanges at a local scale (i.e. upwelling, downwelling, low exchange zones) would control the abundance, taxonomic richness and functional diversity (H3, Dole-Olivier & Marmonier, 1992). Unfavorable conditions for the hyporheic fauna exist in low exchange zones (for both benthic and groundwater fauna); upwelling conditions lead to enhanced taxonomic richness of obligate groundwater-dwellers (stygobites), while downwelling conditions lead to increase the abundance and species richness of both temporary (benthic) and permanent hyporheos.

182

183 2. Study area

184

185 The study was conducted in the lowest 40-km reach of the Ain River, a tributary of the Rhône River, 186 France (Dole-Olivier et al., 2019, Fig. 1). In this sector, the Ain River flows in a large alluvial plain 187 composed of recent alluvia covering 5 to 30 m thick fluvio-glacial deposits, overlying Miocene molasses 188 that represent the impermeable substratum. The alluvial plain width is reduced in its central part by 189 two morainic hills between which the river flows (dotted lines in Fig. 1). In this sector, the mean annual discharge is 120 m³ s⁻¹, with low flow conditions during the summer period (mean July-August, 190 discharge: 54 m³ s⁻¹). The river hydrology is also strongly influenced by five dams located upstream of 191 192 the studied reach that produce daily discharge fluctuations and a significant sediment deficit (Rollet et 193 al., 2014).

194

195 The lower Ain River is relatively wide with a wetted channel width of 51.5 m (± 15.9 m) and an active 196 strip of 89.5 m wide (± 31.2 m, Dole-Olivier et al., 2019). The sediment deficit due to the dams induced 197 a degradation of the riverbed of 2 to 4 cm per year from 1976 to 1999 (Rollet et al., 2014) on the first 198 10 km of the upstream part of the studied sector, around the four upstream gravel bars (noted by a 199 pointed dark line on the Fig. 1). This riverbed degradation resulted in the erosion of fine particles and 200 the local pavement of the riverbed (Rollet et al., 2014; Słowik et al., 2021). From 2005 on, a restoration 201 program progressively took place in this sector of the Ain River, with injection of medium-to-fine 202 sediment, just upstream of gravel bar 5, to reduce the riverbed degradation (Lejot, 2008). A total of 203 89,000 m³ were injected in 6 years (Syndicat de la Rivière d'Ain Aval et de ses Affluents). Presently, the 204 introduced fine sediment covers the river bottom and hides the local pavement (except at gravel bar 205 7).

- 207 3. Material and methods
- 208

206

209 **3.1. Sampling design**

210 Distribution of the hyporheic biodiversity along the Ain River was analysed in 2015 from June 29th 211 to July 3^{rd} when the stream was at its lowest discharge (i.e. between 12.5 and 16 m³ s⁻¹) for at least 10 212 days. The sampling stations were located on gravel bars as they are relevant geomorphological units 213 to predict local downwellings and upwellings (at upstream and downstream positions along the bar respectively, Dole-Olivier and Marmonier, 1992). Of the 40 gravel bars studied in 2014, a total of 15 214 215 gravel bars were selected from Pont d'Ain to Saint Maurice de Gourdans (coded from 5 to 33, similarly 216 to Dole-Olivier et al., 2019; Fig. 1). The gravel bars were sampled at two stations (upstream and 217 downstream of the bar), in three repeated points per position (hereafter called triplicates) randomly 218 chosen 1.5 m apart from one another (Boulton et al., 2003) and at three depths for each point, i.e. -219 20, -40 and -60 cm below the sediment surface (i.e. $15 \times 2 \times 3 \times 3 = 270$ samples). The codes used for 220 stations were labelled by a number for the gravel bar (from 5 to 33) and a letter for the position (U for 221 upstream and D for downstream). These stations were selected to cover most of the studied sector and to include all types of hydrological characteristics: 26 with downwelling or upwelling conditions 222 223 and 4 with low hyporheic exchanges, in a similar proportion to those selected in the study conducted 224 in 2014 by Dole-Olivier et al. (2019) (i.e. 84% of stations with vertical exchanges and 16% with low or 225 no exchanges).

226

227 3.2. Hydrological characteristics of the study sector

228 The hydrogeological characteristics of the Ain alluvial plain and resulting river-aquifer interactions 229 were studied using geomatics analysis (Graillot et al., 2014; Paran and Augeard, 2017; Marmonier et 230 al., 2019). The direction of groundwater/river hydraulic exchanges was inferred from point 231 observations of water levels (or piezometric maps) for both the aquifer and the river. In unconfined 232 alluvial aquifers, where the hydraulic connection with the river is proven, the water flow between the 233 groundwater and the river is proportional to the slope of the free surface of the groundwater near the 234 bank and the permeability of the alluvium following Darcy's law). We consider that the exchanges are 235 made through a vertical plane whose area is defined by the bank length and the water height.

236

237 The exchange evaluation was based on a groundwater model by Burgeap (an environmental 238 engineering group, Costaz, 2005-2006) with the modular three-dimensional finite-difference 239 groundwater flow model Modflow (USGS, Harbaugh et al., 2000). In a second step, the model was 240 improved by D. Mimoun, S. Gaur and F. Paran (Bornette et al., 2014) using river and groundwater 241 levels, volume of water extraction, and hydraulic conductivity obtained from the Syndicat de la Basse Vallée de l'Ain (SBVA), the Centre d'Étude Technique de l'Équipement (CETE Lyon), the Diren Rhône-242 243 Alpes and the French geological survey (BRGM). The resulting regional groundwater flow model (grid 244 size 60 m; 249,000 grid cells) for the Ain basin area (360 km²) was constructed and calibrated for 245 transient-state conditions. The regional piezometric surface was based mainly on water hydrographs 246 for 280 wells in the alluvial plain, while the mean thickness of the surface soil layer and vadose zone is 247 about 3 m. The median values of hydraulic conductivity range from 0.0018 m s⁻¹ (for the old fluvioglacial deposits) to 0.003 m s⁻¹ (for the recent layers). The horizontal hydraulic conductivity has been 248 249 estimated from well tests and published data (Costaz, 2005, 2006) and found to vary between 3.10-5 250 to 4.10-2 m.s-1, with a strong decrease in the central part of the studied sector (between the two 251 morainic hills, downstream of the gravel bars 10 to 12). Specific yield (Ss) values for alluvial deposits 252 ranged from 1 to 17% (mean 5%).

The geomatic analysis was based on the model calibration expressed as a water table contour maps of the alluvial aquifer during a dry period (July 2006). A TIN mesh (Triangulated Irregular Network) was used to build the 3D model of the groundwater surface, using the slope of the water table (in %) and the direction of the flow (in degrees). The permeability of the aquifer (K) was estimated using model calibrated values. The calculation of flow exchanges between the alluvial groundwater and the Ain River was carried out using the ArcGIS Desktop 10.6 software. Because of uncertainties inherent in the geomatic analysis only the direction of exchanges was considered for this study.

260

261 **3.3. Local scale hydrological patterns**

A mobile standpipe was pushed inside the sediment at 20, 40 and 60 cm depth, and 1 L of hyporheic water was pumped using a peristaltic pump. A HACH thermo-conducti-oxymeter was used to measure temperature (° C), electric conductivity (μS cm⁻¹) and dissolved oxygen (mg L⁻¹). Similar measurements were collected concomitantly in the surface water. The vertical hydraulic gradient (VGH) was obtained by measuring the difference between the groundwater level in the mobile piezometer and the surface water level reported for the sampling depth (Baxter et al., 2003).

268

Using conductivity-temperature biplots (Dole-Olivier et al., 2019; Fig. 2A to 2C), the stations were 269 270 separated into three groups according to the direction of the hydrological exchanges: (i) stations 271 located in an upwelling zone with a positive VHG, a low hyporheic temperature (generally below 17°C) and high electric conductivity (generally above 400 µS cm⁻¹, Fig. 2A); (ii) stations located in a 272 273 downwelling zone with negative VHG, low electric conductivity (generally below 400 µS cm⁻¹) and high 274 temperature (generally above 17°C, Fig. 2C) and (iii) stations with weak or no exchange between river 275 and aquifer with null values of VHG combined with low oxygen concentration (average O_2 concentration < 2.5 mg L⁻¹, Fig. 2B). Accordingly, 9 stations were located in "upwelling zones" (Fig. 2A), 276 277 4 stations in "low exchange sites" (Fig. 2B) and 17 stations in "downwelling zones" (Fig. 2C, full data in 278 supplementary material Fig. S1). No upstream-downstream segregation along the study reach was 279 observed in these 3 groups, but a segregation of the stations was found with river geomorphology:

280 most of the downwelling stations (10 out of 17) were located upstream of gravel bars (noted U), 281 whereas most of the upwelling stations (6 out of 9) were located downstream of gravel bars (noted D 282 in the Fig. 2). The low-exchange stations were evenly distributed at the upstream and downstream 283 ends of three different gravel bars.

284

285 3.4. Sediment characteristics

286 Several methods were proposed to estimate the sediment characteristics (Descloux et al., 2010). Yet, 287 most of them only consider the surface layer, despite potential vertical heterogeneity of the river 288 sediment due to variability in fine particle penetration (e.g. Brunke 1999, Gayraud & Philippe, 2001). 289 To evaluate the relevance of these methods based on surficial sediment assessment for hyporheic 290 biodiversity, we used a visual estimation of sediment characteristics based photograph analysis. At 291 each station, thee images of 0.25 m^2 of the surface sediments were randomly taken with a digital 292 camera to define 3 types of substrate, scored as follows (Fig. 2D to 2E): substrate type dominated by 293 sand, gravel and small pebbles (score 1), substrate type defined by a mixture of gravel and large 294 pebbles (score 2), substrate type dominated by large pebbles (score 3). The mean score values 295 (calculated with the three replicate photographs) identified three groups of station (Fig. 2D to 2E): 13 296 stations were characterized by fine sediments, 10 by mixed sediments and 7 by coarse sediments, 297 without upstream-downstream segregation or any relation with the last century eroded zone (stations 298 5 to 9).

299

The sediment stability was estimated using the same images considering biofilm cover (Fig. 2G and 2H). We used differences in biofilm development to evaluate sediment stability because the river discharge was low for only 10 days. The mobile and instable areas were devoid of biofilm (score 0), while the stable sediment was covered by a filamentous biofilm (score 1). The mean score values identified two homogeneous groups: 13 stations were devoid of biofilm, while 17 were covered by filamentous algae. This stability index did not show any upstream-downstream segregation or relation with the last century eroded zone.

307

Coarse Particulate Organic Matter (CPOM) content per mass of dried sediment was measured by loss
on ignition (550°C for 4.5 hours) in the laboratory for each 5L faunal sample (see below) after all
invertebrate removal (CPOM expressed in mg L⁻¹).

311

312 3.5. Hyporheic invertebrates

The hyporheic fauna was collected using Bou-Rouch sampling equipment (Bou & Rouch, 1967,
Stubbington et al., 2016) including a steel standpipe and a piston pump. Each sample consisted of 5 L

315 mixture of water, sediment, particulate organic matter, and organisms. This procedure was applied at 316 each point and depth (-20, -40 and -60 cm). Samples were sieved through a 200 µm mesh after 317 elutriation to retain invertebrates and CPOM, and preserved with 96% alcohol. In the laboratory 318 sorting and identification of invertebrates were performed with the aid of a stereomicroscope 319 Olympus ZSX16 and a light microscope Olympus BX40, when needed. Clitellata, Copepoda, Ostracoda, 320 Cladocera, Amphipoda, and Isopoda were identified to the species level. Flatworms, leeches, molluscs 321 and most insect instars were identified to the genus or species levels (when possible). Diptera were 322 identified to the family or the tribe levels. Finally, Acari Hydrachnidia Hydrachnidae and Nematoda 323 were not identified further.

324

To study spatial patterns of functional diversity, the invertebrates were separated into two functionalgroups:

-Affinity to the groundwater habitat: (i) temporary hyporheos: organisms found inside the sediment that have an obligate aerial stage (i.e. benthic insects, excluding some Coleoptera that have aquatic adults); (ii) permanent hyporheos: organisms without specialization to groundwater life but that can complete their biological cycle without aerial stages (mainly flatworms, leeches, some oligochaetes, molluscs, some micro and macro-crustaceans, and Coleoptera with aquatic adults); and (iii) stygobites: obligate groundwater species that cannot survive in surface water (some oligochaetes, molluscs, and micro and macro-crustaceans in the Ain River).

-Food types used: (i) organisms feeding on biofilm covering fine sediments, hereafter sediment-feeders; (ii) feeding on particulate organic matter, hereafter POM-feeders; (iii) feeding on algae; and (iv) predators. Abundances in each group correspond to the sum of the abundances of all taxa of this group, corrected by a percentage of affinity when a species is known to belong to more than one trophic group. Food type uses were assessed using Tachet et al. (2000) for most benthic organisms, Juget and Lafont (1994) for Clitellata, and Marmonier et al. (1994) and Dole-Olivier et al. (2000) for micro-crustaceans.

341 The functional group diversity was calculated separately on the two types of grouping using 342 the Simpson Index (D = 1 / Σp_i^2) calculated on abundance-weighted functional groups (Gallardo et al., 343 2009; Schmera et al., 2017).

344

345 3.6. Data analysis

The spatial variation in physicochemical characteristics of the hyporheic water (n = 270 samples) was analysed using Principal Component Analysis (PCA) for the 5 parameters (i.e. VHG, electric conductivity, dissolved oxygen (concentration and saturation), and temperature). Concentrations of dissolved oxygen and CPOM were compared among groups of stations defined by sediment grain sizes (fine sediment, mixed, coarse), sediment stability (stable, unstable) or hydrology (upwellings, low
 exchanges, downwellings) using one-way ANOVAs after log-transformation, with mean value per
 station as replicates.

353

At the scale of the study reach, spatial autocorrelation among stations has been estimated for each 354 355 variable using the acf function in R 4.1.2 software (R Core Team, 2021). As these preliminary analyses 356 did not reveal any significant spatial autocorrelation, the longitudinal distribution of the hyporheic 357 fauna was analysed using linear correlation, with the distance downstream of the last dam (expressed 358 in km) as independent variable, while dependent variables were taxonomic richness and abundances 359 of the total assemblages, of the 3 ecological groups and the 4 trophic groups. Taxonomic richness of 360 trophic groups was not used because most taxa belongs to more than one trophic group. The 361 composition of hyporheic assemblages was studied using a Correspondence Analysis (CA, Benzécri, 362 1973) to highlight spatial trends along the study reach and for different groups of stations based on 363 sediment grain size, sediment stability and hydrology. PCA and CA were both performed using 364 ExcelStat 2014. Total taxonomic richness was compared among the three groups based on hydrology. 365 Due to the uneven distribution of samples among downwelling, upwelling and no-exchange stations 366 (153, 81, and 36 samples each), rarefaction curves were generated based on sample-based incidence 367 data to allow comparison of taxa richness at the same level of sampling effort using EstimateS, Version 368 9 (R. K. Colwell, <u>http://purl.oclc.org/estimates</u>).

369

370 At the local scale, statistical differences among taxonomic richness and abundances of all functional 371 groups were investigated for groups of stations based on grain size categories (fine, mixed and coarse), 372 sediment stability categories (stable vs unstable) or hydrology (upwelling, downwelling and low 373 exchange) by one-way ANOVAs with stations as replicates (mean values for all depths and replicates 374 and at the -20cm depth alone). In a last step, abundances and taxonomic richness of the total 375 assemblages, of the ecological groups, and the abundances of the functional groups were compared 376 among depths (-20, -40 and -60 cm depths) and hydrological patterns (downwelling, upwelling, no 377 exchange) using two-way ANOVAs with stations as replicates, after log (x+1) transformation when 378 necessary, followed by a Tukey's test. For all variables, the normality and the homoscedasticity of the 379 residues were verified using the Shapiro-Wilk's test and the Levene's test, respectively. Significance 380 level for statistical tests was set at α < 0.05. All ANOVAs were performed using Statistica (Statsoft Inc., 381 USA).

- 382
- 383 4. Results
- 384

385 4.1. Hyporheic water chemistry and hydrogeology of the sector

386 The dissolved oxygen concentrations in the hyporheic water increased from upstream to downstream 387 $(r^2=0.21, p=0.006; Supplementary material Fig. S3)$, with low oxygen values upstream (stations 5 to 9) and higher values downstream (stations 22 to 33). In contrast, no longitudinal trends for temperature, 388 electric conductivity, VGH and CPOM were detected (Supplementary material Fig. S3). Any longitudinal 389 390 variability for these parameters was better explained by river-groundwater exchanges evaluated by 391 geomatic modelling (Fig. 3A). On one hand, the groundwater flow in the Ain alluvial plain was north-392 east to south-west directed, which resulted in local groundwater inflow along the river (e.g. near the 393 stations 7, 22, 31-33 for the left bank). On the other hand, when the geomatic model was combined 394 with the river forms (here meanders), the hyporheic water chemistry showed repeated inversions of 395 the river-aquifer exchanges between upstream and downstream ends of the meanders (e.g. at gravel 396 bars 12, 15 and at the large meander surrounded by the morainic hills from gravel bars 25 to 29). The 397 geomatic analysis thus supported the majority of the diagnostics based on hyporheic water chemistry 398 (20 of 30 stations, Fig. 3A and B). The stations classified "low exchange" type were poorly predicted by the geomatic analysis (3 of 4 stations were incorrected classified as "groundwater inflow" type by the 399 400 geomatic model). In contrast, most stations classified in the "upwelling zone" type by the hyporheic 401 water chemistry (8 out of 9) were consistent with the geomatic analysis. The only exception was the 402 upwelling observed at station 31U, which occurred in a section where low exchange was predicted by 403 the geomatic analysis (Fig. 3B). In the same way, most of the stations classified as "downwelling zones" 404 by the hyporheic water chemistry (11 of 17) concurred with the geomatic model. The six cases of 405 discrepancy between the geomatic model and the water chemistry were linked to narrow gravel bars 406 located in straight sections of the river (i.e. not associated to meanders). In all cases, the downwelling 407 zones observed at the upstream end of these small gravel bars (i.e. in stations 22U and 29U) or at both 408 ends (i.e. stations 9U-9D and 17U-17D) were not predicted by the geomatic model. In contrast, when 409 the gravel bars were associated with meanders, the prediction based on the geomatic analysis and the 410 hyporheic water chemistry agreed quite well, with a reversion of exchange direction between 411 upstream and downstream positions (i.e. for stations 12U-12D and 15U-15D) or a massive 412 groundwater influence all along the downstream part of the large central meander (i.e. stations 26D, 413 28U-28D, 29D).

414

415 **4.2.** Hyporheic water chemistry and sediment characteristics at the local scale.

416

The hyporheic water characteristics also varied at the station scale, with variations according to the position, the sediment characteristics and the sampling depth. The PCA on the 270 samples (i.e. 15 stations, 2 positions, 3 depths and 3 replicates, Fig. 4) showed a gradient of high electric conductivity
and positive VHG to warmer and well oxygenated water on the 1st axis.

421

422 The consistence of the three groups of stations based on the hydrology (using the biplots of 423 temperature and electric conductivity) could be verified with their location in the PCA (Fig. 4B). The 424 stations classified in upwelling zones (blue dots) had negative coordinates on the 1st axis, stations 425 classified as downwelling zones (red dots) had mostly positive coordinates on the same axis, and 426 stations classified as low exchange (black dots) were all closely grouped on the negative side on the two principal components (Fig. 4B). The distribution of these groups along the 2nd axis was due to a 427 428 significant gradient in dissolved oxygen concentration that decreased from downwelling to upwelling 429 zones and to low exchange areas (Fig. 4E; $F_{(2, 27)}$ = 11.45, p-value = 0.0002). Nevertheless, the three 430 groups of stations were not strictly separated by the PCA: some sampling points pertaining to 431 upwelling and downwelling zones were close to or overlapped with low exchange stations (Fig. 4B). 432 This mixing was due to varying hydrological exchange intensities. For example, the station 12D (fed by 433 groundwater, 16.8°C and 481 μ S cm⁻¹ at -60 cm depth) and the stations 25U (fed by surface water, 434 20.3°C and 385 μ S cm⁻¹ at -60 cm depth) were mixed with "low exchange" stations because of zero 435 VGH. More than three clearly separated groups, the PCA highlighted a gradient of stations with 436 different exchange intensities.

437

In contrast, stations characterized by contrasting sediment grain sizes (Fig. 4C) or sediment stability (Fig. 4D) were poorly or not separated at all by the two first axes of the PCA. The only chemical parameter that significantly changed with grain size was the dissolved oxygen concentration (Fig. 4F; $F_{(2, 27)}=5.21$, p-value = 0.012), with higher mean values below "mixed sediments" than under sediments visually estimated as fine or coarse. Finally, the CPOM concentrations were not significantly different between the groups of stations, regardless of the criteria used (hydrology, grain size or stability) because of high intragroup variability (p-value > 0.05).

445

446 **4.3.** The hyporheic fauna distribution at the sector scale

447

A total of 110,106 individuals belonging to 156 taxa were collected in the 270 samples (see dominant species in the frequency histograms, Supplementary material, Fig. S3). At the scale of the study reach (Fig. 5), both abundances and taxonomic richness of hyporheic fauna increased from upstream to downstream, with very low values measured from gravel bars 5 to 9 and higher values obtained downstream (e.g. gravel bars 30 and 33), resulting in significant correlations with the distance from the last dam located upstream of the studied reach (p-values < 0.01; Fig. 5).

455 One of the ecological groups, the permanent hyporheos, followed a similar increase with distance 456 downstream of the dams (p-values < 0.01; Fig. 5), with very low abundances measured from gravel 457 bars 5 to 9. In contrast, the temporary hyporheos (aquatic insect instars with flying adults) did not 458 show any significant longitudinal trend (p-value > 0.05). High abundances were measured in all 459 stations along the studied sector. The stygobites showed a heterogeneous longitudinal distribution, 460 with high abundances measured in stations located downstream of meanders (i.e. stations 12D and 461 15D), or close to the morainic hills (i.e. gravel bars from 26 to 29) and in two gravel bars downstream 462 of the sector (i.e. 30 and 33). The link between stygobite abundance and the presence of the meanders 463 and morainic hills was associated with stygobite crustaceans, such as the amphipods *Nipharqus* spp. 464 that reached their highest abundances in the stations 12D, 15D, 26D and 28U-D (Fig. 5), four stations 465 located in areas where the hydrological model predicted groundwater inflows (Fig. 3).

466

467 All trophic functional groups followed a significant increase with the distance downstream of dams (p-468 values < 0.01; Fig. 5). The upstream area harbored low abundances of all feeding groups, which 469 increased downstream. When the proportions (in %) were calculated, the sediment feeders were 470 dominant in the upstream part of the sector (more than 50% of the individuals) and decreased 471 downstream (negative correlation with distance $r^2 = 0.270$, p-value = 0.0016), while the POM feeders 472 were poorly represented in upstream areas (around 5% of the assemblage) and increased with the distance downstream ($r^2 = 0.274$, p-value = 0.0015). No significant trend was observed in the 473 474 percentages of algae feeders and predators at the scale of the study reach.

475

476 4.4. Effect of sediment characteristics and local hydrology on hyporheic fauna at the station scale 477

478 The Correspondance Analysis (Fig. 6) highlighted a gradient in the composition of the assemblages, but 479 with a rather low percentage of variance explained by the two first axes (11.3%). The 1st axis 480 represented a gradient of taxa with different affinities to groundwater. Most stygobite species were 481 located on the negative side of this axis: oligochaetes Haber turquini, Trichodrilus spp., Rhyacodrilus 482 balmensis, Spiridion phreaticola, the mollusc Spiralix sp., Bythiospeum cf diaphanum, Islamia cf minuta, 483 the Crustacea Microcharon reginae, Parabathynella cf stygia, Salentinella angelieri, Niphargus spp., Proasellus sp., Parastenocaris cf glareola, Phreatalona phreatica, Cryptocandona kieferi and 484 485 Marmocandona zschokkei. On the positive side of this axis, most species were burrowers in fine 486 sediments (e.g. the Clitellata Lophochaeta ignota, Tubifex tubifex, Tubifex spp., Potamothrix 487 moldaviensis, the Epheremoptera Ephemera sp., the Diptera Chironomini and Tanytarsini) together 488 with species living at the surface in moderate to slow flowing zones of rivers (e.g. Erpobdella

octoculata, Glossiphonia sp., *Asellus aquaticus, Cypria ophtalmica*). Finally, a group of taxa were
located close to the origin of the 1st axis, but with negative values on the 2nd axis, largely consisting of
insects (e.g. *Potamanthus luteus, Micronecta* sp., *Stenelmis* sp., *Leuctra* cf *nigra, Corinoneura* sp.) or
epigean crustaceans (e.g. *Alona rectangula, Chydorus sphaericus, Cypridopsis vidua*) that mostly live in
the benthic layer of rivers. As some stygobite species have positive coordinates on the 2nd axis (e.g. *Spiralix* sp., *Microcharon* cf *reginae* or *Parabathynella* cf *stygia*) there is a light redundancy between
the 1st and the 2nd axis that induced an arc distribution of the samples (Fig. 6D to F).

Considering the distribution of samples along the two first axes (Fig. 6D to F), sediment characteristics 496 (grain size (Fig. 6E) and stability (Fig. 6F)) had poor influences on the hyporheic fauna: samples of the 497 498 different groups strongly overlapped. Similarly, no differences were observed for the abundance and 499 taxonomic richness of the whole assemblage, for the ecological groups and for the trophic functional 500 groups under coarse, mixed, and fine sediment or under stable or unstable sediment patches (Table 1, 501 Supplementary material Fig. S4, in all cases p-values < 0.05). This lack of difference was also observed 502 when the -20 cm depth was analysed alone (p-values < 0.05 for abundances and richness of all taxa 503 groups).

504

505 In contrast, the local hydrology influenced the composition of the hyporheic assemblages (Fig. 6D). The 506 samples from upwelling zones were all located on the negative side of the 1st axis, closely associated 507 with the stygobite fauna. A part of the samples from the downwelling and most of those from low 508 exchange zones were mixed together on the positive side of the 1st axis, closely associated with species 509 living in slow flowing water and fine sediment. Finally, a large set of samples from downwelling zones was located close to the origin of the 1st axis and on the negative side of the 2nd axis, closely associated 510 511 to benthic species. These differences in assemblage composition among hydrology groups were also 512 linked to higher abundances and taxonomic richness in areas with vertical exchanges (i.e. upwelling 513 and downwelling zones) than in low exchange areas. The total abundance, the abundance of the 514 temporary hyporheos and the abundances of organisms feeding on fine sediment, POM and algae 515 were significantly higher in the stations located in downwelling zones than in the two other contexts 516 (Table 1, Supplementary material Fig. S4). The stygobites followed a rather different trend with higher 517 abundance and richness found in stations located in upwelling zones (but only significant for taxonomic richness, Table 1, see stygobite abundance in the Supplementary material Fig. S4). 518

519

520 **4.5.** Local hydrology, taxonomic richness and functional groups.

521 The local hydrology (downwelling, upwelling and low exchange) had a strong effect on the taxonomic 522 richness of the hyporheic assemblages. The taxonomic richness estimated by rarefaction curves (i.e. for a similar sampling effort of 36 samples, Fig. 7) was consistently higher in stations located in the downwelling and upwelling zones than in stations from low exchange zones.

525

In addition to local hydrology, the sampling depths had major effects on the hyporheic assemblages 526 527 (Fig. 8): the abundances and the taxonomic richness of 1) all fauna, 2) the temporary and permanent 528 hyporheos and 3) the abundances of all food type groups decreased from the downwelling zones to 529 the low exchange areas and with depths (from -20 to -60 cm depths). These changes linked to 530 hydrology were similar at all depths (no significant interaction between hydrology and depth was 531 detected, see Supplementary material Table S2). A very different pattern was observed for stygobites 532 (Fig. 8). These organisms adapted to groundwater habitats reached their highest species richness and 533 abundances in the upwelling zones and did not show any significant change with depth.

534 In addition, the total taxonomic richness and total abundances did not uniquely differ with the 535 direction of water exchanges, but also changed with the intensity of these exchanges. This was particularly clear at -60 cm depth in the downwelling zones, where the taxonomic richness and the 536 abundance increased with the water temperature ($r^2=0.159^{**}$ and 0.155^{**} for taxonomic richness and 537 538 total abundance, respectively) and with the concentration in dissolved oxygen ($r^2=0.257^{***}$ and 539 0.158** for taxonomic richness and total abundance, respectively, Supplementary material, Fig. S5) is 540 highlighting a more abundant and diversified hyporheic fauna in areas where the infiltration of surface 541 water was most intense.

542

543 **4.6. Functional diversity**

Different trends were observed for the two types of functional groups (Fig. 9). The functional diversity based on food type groups significantly increased with the distance downstream of the last dam $(r^2=0.31, p-value = 0.0006)$. In contrast, the functional diversity based on the ecological groups (i.e. affinities with groundwater) did not increase as predicted from upstream to downstream. Functional diversity reached maximum values in the central part of the studied sector (resulting in a correlation with a bell-shaped trend, $r^2=0.21$, p-value = 0.0052), but with high values in two downstream stations (29D and 31U), located in upwelling zones.

551

The functional diversity based on ecological groups was higher in the upwelling zones compared to low exchange areas (without vertical trend; Fig. 9), while the functional diversity based on food type groups changed with the combination of local hydrology and depth. The later was slightly higher in the two groups of stations with vertical exchanges compared to low exchange zones, and decreased significantly with depth (at least in downwelling and upwelling zones).

558 The diversity based on affinity to groundwater was mainly controlled by the relative abundance of 559 stygobites in the assemblage. The diversity index was significantly correlated to the percentage of stygobites for the longitudinal trend (r^2 =0.13, p-value = 0.022) and for the local hydrology (r^2 = 0.59, p-560 value = 0.03). The diversity based on food type groups increased with the decreasing relative 561 abundance of sediment feeders (negative correlation $r^2=0.95$, p-value = 4.10⁻²⁰), the increasing relative 562 abundances of predators (r^2 =0.45, p-value = 2.10⁻⁵) and POM feeders (r^2 =0.37, p-value = 0.0002), two 563 564 groups that followed the same upstream-downstream increase in the hyporheos. A similar relationship 565 with the relative abundances of sediment feeders (negative correlation $r^2=0.86$, p-value = 0.003) was 566 observed when the depth and the hydrology were combined.

567

568 5. Discussion

569

570 **5.1.** Distribution of the hyporheic fauna at the scale of the study reach.

The hypothesis of a combined control of hyporheic fauna distribution by hydrogeology of the valley and geomorphology of the river channel (**H1**) was clearly verified when considering the distribution of the hyporheic fauna along the studied 40-km reach. Several studies highlight the role of large scale hydrogeological heterogeneity and local scale geomorphology in the spatial patterns of rivergroundwater exchanges (e.g. Faulkner et al., 2012, Gomez-Velez et al., 2017), especially the location of large-scale draining sectors (i.e. sections of several kilometers where local upwelling zones are fed by deep groundwater, Marmonier et al., 2020).

578

579 In the Ain River, these large-scale groundwater inflows were mostly located in the central part of the 580 studied sector, controlled by the dominant direction of the groundwater flow (illustrated by the 581 geomatic model) and by the presence of the two morainic hills. At this regional scale, the groundwater 582 inflows had a weak influence on the richness and abundances of the hyporheic assemblages, of the permanent and the temporary hyporheos and on the abundance of the trophic functional groups. 583 584 However, we found a significant effect on the stygobite abundance and taxonomic richness. The 585 stygobites can establish permanent and sometime abundant populations at shallow depths, in areas 586 where the environmental characteristics (e.g. temperature, solute contents, flow velocity) exhibit low 587 temporal variability (Dole-Olivier and Marmonier, 1992; Malard et al., 2003). For example, several 588 Niphargus species (here N. casparyi, N. rhenorhodanensis and N. fontanus) are known to live in the 589 hyporheic zone of other tributaries of the Rhône River, especially in upwelling areas of gaining sectors 590 (Marmonier et al., 2019, 2020). In the Ain River, they were mostly sampled downstream of meanders 591 located in the central part of the sector (station 12D and 15D) or close to the morainic hills (stations 592 26D, 28U-D). In addition to these very abundant amphipods, some other rare stygobite crustaceans 593 (e.g. Parabathynella cf stygia and Microcharon reginae according to Dole and Chessel, 1986; Dole-594 Olivier et al., 2009) were sampled here, representing shallow populations of species living deeper in 595 the sediments. These species are rare (in abundance and distribution) and represent a high heritage 596 value for the region. Thus, they must be considered in the management plan for the protection of 597 river-groundwater exchanges and global river biodiversity (Michel et al, 2009; Boulton, 2020). The 598 proportion of stygobites in the assemblages control the level of functional diversity (based on the 599 ecological groups). In addition to their importance in the total taxonomic richness (e.g. Dole-Olivier et 600 al., 2015; Marmonier et al., 2020), their role in ecological functions must be considered with attention 601 (Ercoli et al., 2019; Fattorini et al., 2020; Di Lorenzo et al., 2021).

602

603 This study also highlights the importance of the geomorphology of the Ain River. The shape and 604 location of the channel within its floodplain, and the occurrence of meanders are essential for river-605 groundwater exchanges (Magliozzi et al., 2018), because these channel forms induce intra-meander 606 hyporheic flows (Boano et al., 2006). The hydrological role of meanders has been documented by 607 several authors, both for water exchanges between the river and the hyporheic zone (Stonedahl et al., 608 2013; Nelson et al., 2019) and for biogeochemical processes (e.g. for nitrogen dynamics, Dwivedi et 609 al., 2018). In the present study, all gravel bars located along meanders showed clear downwelling-610 upwelling successions (e.g. stations 12U-D, 15U-D, 25U to 28D). In contrast, gravel bars in straight 611 sections were generally narrow and did not follow the expectation of the geomatic modelling. For 612 example, stations 22U and 29U, predicted to be in two upwelling zones by the geomatic model, were 613 found in downwelling conditions (based on water chemistry). These narrow and straight lateral bars 614 were not taken into account by the large-scale geomatic model, even if they were large enough to 615 support downwelling conditions at their head and upwelling at their tail (Dole-Olivier & Marmonier, 616 1992). Thus large-scale hydrogeological modelling has some limits for integrating small gravel bars and 617 local heterogeneities in the prediction of hydrological exchanges (Stonedahl et al., 2013).

618

619 Finally, the history of river incision was surprisingly important for most of the characteristics of the 620 hyporheic fauna at the 40-km scale. Abundances and taxonomic richness of the total assemblages, and 621 of the permanent hyporheos and the abundances of all food type groups increased with the distance downstream, because of very low values in the four upstream gravel bars (from 5 to 9). These stations 622 623 were located in an area where an intense incision occurred during the XXth century, after the 624 construction of dams and the reduction of sediment transit (Rollet et al., 2014). This was particularly 625 surprising because the riverbed erosion is not yet visible. The recent restoration program with 626 sediment reinjection in the main channel (Lejot, 2008) resulted in fine sediment deposits at the surface 627 (e.g. at gravel bars 5, 8 and 9). Nevertheless, the historical erosion, while completely hidden, is still 628 detectable in the composition of the hyporheic fauna. During river incision, the erosion of sand and 629 gravel generated a re-arrangement of coarse pebbles resulting in a pavement of the river bottom and 630 an accumulation of fine sediment between these large particles (Gomez, 1984; Jain, 1990). In these 631 conditions, the habitability of the interstices progressively decreased and apparently still persisted 10 632 years after the beginning of the injection of gravel and sand in the main channel. The effect of the 633 historical pavement of the upstream sector was also visible in the low concentrations of dissolved 634 oxygen (i.e. positive correlation with distance to the dams), generally associated with low vertical 635 exchanges (Lefebvre et al., 2005) and the reduction of the abundances and richness of the hyporheic 636 fauna (Strayer et al., 1997). None of the ecological groups were affected in a similar way: the very 637 mobile temporary hyporheos (i.e. the aquatic insect instars with flying adults) did not exhibit its lowest 638 richness and abundances in the upstream stations. While the less mobile permanent hyporheos (i.e. 639 Clitellata, Mollusca and Crustacea) showed lower abundance and lower richness in the four upstream 640 gravel bars than downstream. In addition, the hyporheic assemblages sampled in these stations 641 showed specific feeding traits, with a dominance of the fine sediment feeders, certainly favored here 642 by the storage of fine particles. This dominance resulted in lower functional diversity, which increased 643 downstream together with the proportion of other trophic groups, such as POM feeders. Some of 644 these POM feeders, being attracted by porous and mobile sediment (e.g. the Gammarus species, 645 Vadher et al., 2015; Navel et al., 2010), were more abundant downstream. The functional diversity 646 based on trophic traits appeared thus to be sensitive to the physical characteristics of the hyporheic 647 zone, even if differences are not visually obvious.

648

649 **5.2. Weak influence of surface grain size and sediment stability**

650 Surprisingly, our results did not support the influence of heterogeneity in surface sediment grain size 651 and sediment stability on the hyporheic fauna (H2 rejected). The importance of sediment grain size 652 and heterogeneity on hyporheic processes has been widely documented (Richards and Bacon, 1994; 653 Wood and Armitage, 1997; Tonina et al., 2016; Nelson et al., 2019). For example, decreases in dissolved 654 oxygen concentrations and nitrification processes were generally observed in fine sediment, while 655 ammonium concentrations and denitrification process increased (Dahm et al., 1987; Lefebvre et al., 656 2005). This shift from oxic to hypoxic or anoxic processes was related to fine sediment being the 657 substrate of the biofilm, which stimulated its growth and respiration (Claret and Fontvieille, 1997) and 658 decreased vertical hydrologic exchanges and oxygen supply in the hyporheic zone (Tonina and 659 Buffington, 2009; Cook et al., 2020). A relationship between surface sediment grain size and hyporheic 660 processes was not always detected (Descloux et al., 2010; Nogaro et al., 2010), but appeared strong 661 enough to justify methods of visual assessment of surface sediment characteristics for river bed 662 clogging (Peck et al., 2000; Sennatt et al., 2006). In the present study, very few changes were observed

in the habitat characteristics and in the fauna composition between areas differing in surface sediment 663 grain size. First, the hyporheic water chemistry was poorly controlled by the surface sediment grain 664 665 size. Most chemical parameters did not change at all, and only a minor increase in dissolved oxygen 666 concentrations was measured at stations with mixed sediment size and lower values in the fine and 667 coarse sediment contexts. This increase in oxygen content in mixed sediment was probably a side effect of the local hydrology: stations 22U, 30D and 33D were all located in strong downwelling zones, 668 with more than 11 mg L⁻¹ of dissolved oxygen at 60 cm depth inside sediments. Second, the expected 669 decrease in the hyporheos abundance and taxonomic richness with decreasing sediment size was not 670 671 observed in this study: the surface characteristics of the bed sediment did not explain the composition 672 and the functional diversity of the hyporheic assemblages. This was likely due to a strong vertical 673 stratification of bed sediment (as observed with a geological radar by Mermillod-Blondin et al., 2015 674 or with field measures and modelling by Cardenas and Zlotnik, 2003) that generally controls hyporheic 675 water fluxes. The vertical stratification of river sediment is linked to the history of the sediment origin, 676 transit and deposition (Marion et al., 2008). The visual estimate of surface sediment grain size may be 677 efficient for small streams with a thin layer of bottom sediment, but it is less informative for a 678 consistent diagnostic of hyporheic habitat quality in large gravel-bed rivers (Nogaro et al., 2010; 679 Descloux et al., 2010).

680

681 The stability or mobility of surface sediment has also been evaluated as an explanation for changes in 682 hyporheic processes (e.g. Peterson et al., 2008; Zheng et al., 2019; Dudunake et al., 2020). Sediment 683 mobility may have different effect on hyporheic biodiversity because of direct (invertebrate migration) 684 and indirect consequences (sediment modifications). At a small time-scale sediment mobility may 685 induce an increase in invertebrate drift (Gibbins et al., 2007) and in the vertical migration of the 686 hyporheic fauna (Dole-Olivier et al., 1997, Schmid-Araya, 2000). At a long time-scale, a low mobility of 687 bed sediment may reduce the resuspension of fine particles, resulting in local clogging of the interstices, and the formation of anoxic microzones with local denitrification (Roy Chowdhury et al., 688 689 2020). In the studied sector, the development of a filamentous biofilm at the surface of the particles 690 was not a consistent predictor of the hyporheic habitat suitability and hyporheos composition. Even 691 the expected increase in algae-feeder densities below the thick biofilm was not supported, probably 692 because of very different porosity under similar level of biofilm development. Hence, the use of surface 693 biofilm development to evaluate sediment stability seems poorly informative. In the future studies, 694 the sediment stability must be considered using combined methods, such as hydraulic modelling (e.g. 695 Buffington and Montgomery, 1997) or direct measures of sediment movement (e.g. spray-painted or 696 magnetically tagged stones, Mao and Surian, 2010) to evaluate gravel mobility.

698 **5.3. Local hydrology, the major driver for hyporheic biodiversity**

The present study clearly demonstrated that local hydrology (i.e. downwelling, upwelling *versus* low exchange areas) had a major role in the distribution and composition of hyporheic fauna (**H3** verified). Several significant differences were observed in the abundance, the taxonomic richness and the functional diversity according to the hydrological connections, with a similar trend at all depths.

703

704 First, specific environmental and faunal characteristics were observed in low exchange areas. Here the 705 zero VHG suggested nearly immobile hyporheic water with poor exchanges with both surface water 706 and groundwater. These hydrological conditions resulted in very low oxygen concentrations (i.e. less 707 than 2 mg.L⁻¹ at -60 cm depth). Similar low values in dissolved oxygen were measured in poorly 708 connected riverbeds and were generally associated with denitrification and ammonium accumulation 709 (Dahm et al., 1987; Lefebvre et al., 2005; Marzadri et al., 2012; Harvey et al., 2013). In this study, the 710 low water exchanges induced low hyporheic abundances and low taxonomic richness (for total 711 hyporheos, most ecological groups and the food type groups) compared to the well-connected stations 712 located in downwelling or upwelling zones. The negative impact of a reduction in the water exchange 713 on the hyporheic fauna has been well documented (Brunke and Gonser, 1999; Hunt and Stanley, 2003; 714 O'Sullivan et al., 2019). The hyporheos abundance was generally correlated positively with dissolved 715 oxygen concentrations (Strayer et al., 1997; Boon et al., 2016) and negatively with the accumulation 716 of potentially toxic ammonia inside sediment (Dahm et al., 1987; Dehedin et al., 2013; Caschetto et al., 717 2014, 2017; Di Lorenzo et al., 2021). In contrast, the consequence of a reduced hydrological vertical 718 exchange on the functional diversity is less understood. In this study, the functional diversity evaluated 719 by the ecological groups (i.e. the affinity of organisms with groundwater) followed a similar pattern as 720 taxonomic richness with lower values in the low exchange area than in the other hydrological contexts. 721 While the low taxonomic richness was due to the reduction in the number of temporary hyporheic 722 species (i.e. the benthic insects), the low values in functional diversity were linked to a lower 723 proportion of stygobites. The limitation of vertical exchanges reduced the hyporheic assemblages to a 724 set of permanent hyporheic species (mostly oligochaetes and micro-crustaceans) as already observed 725 in sandy and lowland rivers (e.g. Palmer et al., 1992).

726

In areas fed by groundwater (i.e. upwelling zones), the environmental conditions were less stressful for the hyporheic fauna because of rather large interstices (fine sediment washout) and regular water renewal. These conditions resulted in intermediate dissolved oxygen concentrations (at least in the studied sector where groundwater-fed springs contains between 5.2 and 8.7 mg.L⁻¹ of oxygen, Dehédin, 2012). Here, the total hyporheic abundance was intermediate between the low exchange zones and the downwelling zones. In contrast, the taxonomic richness was high in upwelling zones, 733 because of the large number of stygobite species. Such increases in stygobite richness and abundance 734 in upwellings were frequently observed in a wide range of rivers (Dole-Olivier and Marmonier, 1992; 735 Malard et al., 2003; Di Lorenzo et al., 2013). The upwellings did not represent a real storage zone for 736 river biomass (Dole-Olivier, 2011), but they appeared to be hotspots for biodiversity, especially for rare 737 stygobite species. The high relative abundance of stygobites in the upwelling zones induced an increase 738 in functional diversity based on ecological groups, which did not decrease with increasing depth, 739 because of the ability of most of the ambient stygobites to live in deep river sediment (Dole and 740 Chessel, 1986). In contrast, the functional diversity based on food types (that poorly changed with the 741 local hydrology) decreased with depth in the upwelling zones. This vertical pattern was probably due 742 to a reduction of food diversity with depth, selecting preferentially sediment feeders, which was the 743 trophic group feeding on the most available resource: fine sediments and associated biofilm.

744

745 Finally, in the downwelling zones, the abundance and taxonomic richness were the highest, especially 746 for the temporary hyporheos (i.e. benthic insects with flying adults). This high density may be explained 747 by the inflow of surface water into the sediment, which brings dissolved oxygen and organic matter 748 (Brunke & Gonser, 1999; Franken et al., 2001). This study demonstrates that downwelling zones 749 represent essential storage zones for the benthic fauna, contributing to river resilience through 750 recolonization process of the benthic layer (Dole-Olivier, 2011, Vander Vorste et al., 2016). Although 751 not all downwelling zones were similarly populated: the highest densities of the temporary hyporheos 752 were measured in strong downwellings. This influence of infiltration intensity was illustrated by the 753 positive correlation between the abundance and taxonomic richness with temperature and dissolved 754 oxygen at -60 cm depth. Intense downwellings represented attractive areas for faunal vertical 755 migration (Dole-Olivier, 2011). This richness in benthic species was probably due to the higher diversity 756 of food types in the downwelling zones than in other zones (upwelling and no exchange zones). These 757 downwelling areas had high dissolved oxygen concentration and fresh organic matter, which was 758 reflected in a higher proportion of POM feeders, algae feeders and predators in comparison with other 759 zones with no direct supply of fresh organic matter from surface waters. Therefore, the downwellings 760 were hotspots for biomass and taxonomic richness in the hyporheic zone, but with a different 761 assemblage of species compared to upwelling zones. The importance of upwelling zones is often 762 emphasized by river managers, particularly because of their role in fish reproduction and survival 763 during low water periods (e.g. Curry and Noakes, 1995; Malcolm et al., 2005). In contrast, the 764 importance of downwelling zones is rarely considered. This study supports the idea that the 765 heterogeneity of hydrological patterns, with alternation of upwellings and downwellings, may 766 represent the optimal spatial structure for biodiversity conservation in rivers, with both contributing 767 to the diversity and resilience of the river (Leigh et al., 2016).

769 6. Conclusions

770

771 At the local scale, the composition of the hyporheos was controlled by vertical exchanges of water 772 between the river, the hyporheic zone and the deep groundwater (i.e. downwelling, upwelling or low 773 exchange areas). The upwellings were hotspot of taxonomic richness, but also for functional diversity 774 based on ecological groups. Here, the functional diversity was mostly related to the relative abundance 775 of stygobites and the reduction of dominance of the permanent hyporheos. In contrast, the 776 downwelling zones were hotspots of abundances, especially for the temporary hyporheos (mostly 777 benthic insects). These organisms, living temporarily inside sediment, contribute to the river biomass 778 and resilience. In downwellings, the food-type diversity was mostly controlled by the proportion of 779 POM feeders and the reduction in dominance of sediment feeders. Low exchange zones appeared to 780 be less populated areas with low functional diversity.

781

Compared to hydrology, the surface characteristics of the riverbed sediment (i.e. visual estimation of
the grain size and the stability) was not informative for the hyporheos composition of this large gravelbed river. The visual estimation of surface sediment characteristics neglected the vertical stratification
of the sediment, which is a consequence of the history of sediment transit and deposition.

786

787 These links between vertical water exchanges and hyporheic assemblages must be combined with 788 large-scale patterns of hyporheic distribution. In the 40-km long reach assessed in this study, the 789 hyporheic fauna was controlled by a combination of two factors: the hydrogeology of the alluvial plain 790 (i.e. large-scale groundwater movement) and the geomorphology of the river channel (i.e. location of 791 meanders and history of the river incision). In consequence, river managers must carefully consider 792 the ongoing activities that modify the dynamics of the river-groundwater exchanges (e.g. river 793 regulation or groundwater extraction), but also the long-term history of sediment fluxes due to past 794 regulations.

795

796 **CRediT authorship contribution statement**.

M.J. Dole-Olivier: Conceptualization, Investigation, Writing – original draft. M. Creuzé des Châtelliers:
Conceptualization, Investigation. D. Galassi: Investigation. M. Lafont: Investigation. F. MermillodBlondin: Investigation. F. Paran: Investigation. D. Graillot: Investigation. S. Gaur: Investigation. P.
Marmonier: Conceptualization, Investigation, Writing – original draft.

801

802 **Declaration of competing interest**

- 803 The authors declare the following financial interests, which may be considered as potential competing
- 804 interests: Electricité de France (EDF) who built and manage the upstream dams funded the research
- 805 program, however the LTSER Rhône River Basin and the Rhône River Water Agency were included in
- the following committee and guaranty the objectivity of our conclusions.
- 807
- 808

809 References.

- 810 Baxter, C., Hauer, F. R., Woessner, W. W. 2003. Measuring groundwater–stream water exchange: new
- 811 techniques for installing minipiezometers and estimating hydraulic conductivity. Transactions of
- 812
 the American Fisheries Society.
 132(3),
 493-502.
 https://doi.org/10.1577/1548

 813
 8659(2003)132<0493:MGWENT>2.0.CO;2
- Benzécri, J. P. 1973. L'Analyse des Données. Volume II. L'Analyse des Correspondances. Dunod Ed.,
 Paris.
- Boano, F., Camporeale, C., Revelli, R., Ridolfi, L. 2006. Sinuosity-driven hyporheic exchange in
 meandering rivers. Geophysical Research Letters. 33(18), 1-4. <u>https://doi.org/10.1029/</u>
 2006GL027630
- 819 Boon, P. J., Willby, N., Gilvear, D., Pryce, D. 2016. The regional hyporheic fauna of gravel-bed rivers and
- environmental controls on its distribution. Fundamental and Applied Limnology, 187(3), 223-239.
 https://doi.org/10.1127/fal/2016/0705
- Bornette, G., Foulquier, A., Graillot, D., Marmonier, P., Mimoun, D., Montuelle, B. 2014. Biodiversité
 et fonctions des zones humides en réponse aux étiages sévères induits par le changement global.
- 824 Université Lyon 1, Mines Saint-Etienne, Cemagref Lyon, Projet ANR-CEP09_446043 Wetchange,
 825 Final Report.
- Bou, C., Rouch, R., 1967. Un nouveau champ de recherches sur la faune aquatique souterraine.
 Compte-Rendu Académie des Sciences, 265, 369-370.
- Boulton, A. J., Findlay, S., Marmonier, P., Stanley, E. H., Valett, H. M. 1998. The functional significance
 of the hyporheic zone in streams and rivers. Annual review of Ecology and systematics, 29(1), 5981. https://doi.org/10.1146/annurev.ecolsys.29.1.59
- 831 Boulton, A. J. 2020. Conservation of groundwaters and their dependent ecosystems: Integrating 832 molecular taxonomy, systematic reserve planning and cultural values. Aquatic Conservation:
- 833 Marine and Freshwater Ecosystems, 30(1), 1-7. <u>https://doi.org/10.1002/aqc.3268</u>
- Brown, L. E., Khamis, K., Wilkes, M., Blaen, P., Brittain, J. E., Carrivick, J. L., Fel, S., Friberg, N., Fureder,
 L., Gislason, G.M., Hainie, S., Hannah, D.M., James W.H.M., Lencioni, V., Olafson, J.S., Robinson,
- 836 C.T., Saltveit, S.J., Thomlpson, C., Milner, A. M. 2018. Functional diversity and community assembly
- 837 of river invertebrates show globally consistent responses to decreasing glacier cover. Nature
- 838 Ecology & Evolution, 2(2), 325-333. <u>https://doi.org/10.1038/s41559-017-0426-x</u>
- Brunke, M. 1999. Colmation and depth filtration within streambeds: retention of particles in hyporheic
 interstices. International Review of Hydrobiology, 84(2), 99-117. <u>https://doi.org</u>
 /10.1002/iroh.199900014

- 842 Brunke, M., Gonser, T. 1999. Hyporheic invertebrates: the clinal nature of interstitial communities 843 structured by hydrological exchange and environmental gradients. Journal of the North American
- 844 Benthological Society, *18*(3), 344-362. <u>https://doi.org/10.2307/1468448</u>
- 845 Buffington, J. M., Montgomery, D. R. 1997. A systematic analysis of eight decades of incipient motion
- studies, with special reference to gravel-bedded rivers. Water Resources Research, 33(8), 19932029. <u>https://doi.org/10.1029/96WR03190</u>
- Buffington, J. M., Tonina, D. 2009. Hyporheic exchange in mountain rivers II: Effects of channel
 morphology on mechanics, scales, and rates of exchange. Geography Compass, *3*(3), 1038-1062.
 https://doi.org/10.1111/j.1749-8198.2009.00225.x
- 851 Capderrey, C., Datry, T., Foulquier, A., Claret, C., Malard, F. 2013. Invertebrate distribution across
- nested geomorphic features in braided-river landscapes. Freshwater Science, 32(4), 1188-1204.
 https://doi.org/10.1899/12-188.1
- Cardenas, M. B., Zlotnik, V. A. 2003. Three-dimensional model of modern channel bend deposits.
 Water Resources Research, 39(6), 1-12. <u>https://doi.org/10.1029/2002WR001383</u>
- Cardenas, M. B., Wilson, J. L., Zlotnik, V. A. 2004. Impact of heterogeneity, bed forms, and stream
 curvature on subchannel hyporheic exchange. Water Resources Research, 40(8), 1-13.
 https://doi.org/10.1029/2004WR003008
- Caschetto, M., Barbieri, M., Galassi, D.M.P., Mastrollio, L., Rusi, S., Stoch, F., Di Cioccio A., Petita, M.
 2014. Human alteration of groundwater–surface water interactions (Sagittario River, Central Italy):
 implication for flow regime, contaminant fate and invertebrate response. Environmental Earth
 Sciences, 71, 1791–1807. <u>https://doi.org/10.1007/s12665-013-2584-8</u>
- Caschetto M., Galassi D.M.P., Petitta M., Aravena R. 2017. Evaluation of the sources of nitrogen
 compounds and their influence on the biological communities in the hyporheic zone of the
 Sagittario River, Italy: an isotopic and biological approach. *Italian Journal of Geosciences* 136(2),
- 866 145-156. <u>https://doi.org/10.3301/IJG.2016.07</u>
- Claret, C., Fontvieille, D. 1997. Characteristics of biofilm assemblages in two contrasted hydrodynamic
 and trophic contexts. Microbial Ecology, 34(1), 49-57. <u>https://doi.org/10.1007/s002489900033</u>
- 869 Claret, C., Marmonier, P., Bravard, J. P. 1998. Seasonal dynamics of nutrient and biofilm in interstitial
- habitats of two contrasting riffles in a regulated large river. Aquatic Sciences, 60(1), 33-55.
 https://doi.org/10.1007/s000270050025
- Cook, S., Price, O., King, A., Finnegan, C., van Egmond, R., Schäfer, H., Pearson, J.M., Abolfathi, S.,
 Bending, G. D. 2020. Bedform characteristics and biofilm community development interact to
 modify hyporheic exchange. Science of the Total Environment, 749, 141397.
 https://doi.org/10.1016/j.scitotenv.2020.141397

- 876 Cornut, J., Elger, A., Lambrigot, D., Marmonier, P., Chauvet, E. 2010. Early stages of leaf decomposition
- are mediated by aquatic fungi in the hyporheic zone of woodland streams. Freshwater Biology,
- 878 55(12), 2541-2556. <u>https://doi.org/10.1111/j.1365-2427.2010.02483.x</u>
- Costaz, I. 2005-2006. Modélisation de la nappe alluviale de la basse plaine de l'Ain et de ses milieux
 annexes. Burgeap, Conseil Général de l'Ain, Rapports phases 1 (27p.), 2 (23p.) et 3 (21p.).
- Crenshaw, C. L., Valett, H. M., Tank, J. L. 2002. Effects of coarse particulate organic matter on fungal
 biomass and invertebrate density in the subsurface of a headwater stream. Journal of the North
 American Benthological Society, 21(1), 28-42. https://doi.org/10.2307/1468297
- Curry, R. A., Noakes, D. L. 1995. Groundwater and the selection of spawning sites by brook trout
 (*Salvelinus fontinalis*). Canadian Journal of Fisheries and Aquatic Sciences, 52(8), 1733-1740.
 https://doi.org/10.1139/f95-765
- Dahm, C. N., Trotter, E. H., Sedell, J. R. 1987. Role of anaerobic zones and processes in stream
 ecosystem productivity, in Averett R.C., McKnight D.M. (Eds.), Chemical Quality of Water and the
 Hydrologic Cycle, Lewis Publishers, Chelsea, pp 157-178.
- Datry, T. 2012. Benthic and hyporheic invertebrate assemblages along a flow intermittence gradient:
 effects of duration of dry events. Freshwater Biology, 57(3), 563-574.
 https://doi.org/10.1111/j.1365-2427.2011.02725.x
- Dehedin, A. 2012. Changements globaux et assèchement des zones humides fluviales : conséquences
 sur les processus biogéochimiques et les communautés d'invertébrés. PhD Thesis, University of
 Lyon 1, 254p.
- 896Dehedin, A., Piscart, C., Marmonier, P. 2013. Seasonal variations of the effect of temperature on lethal
- and sublethal toxicities of ammonia for three common freshwater shredders. Chemosphere, 90(3),
 1016-1022. <u>https://doi.org/10.1016/j.chemosphere.2012.07.055</u>
- Descloux, S., Datry, T., Philippe, M., & Marmonier, P. 2010. Comparison of different techniques to
 assess surface and subsurface streambed colmation with fine sediments. International Review of
 Hydrobiology, 95(6), 520-540. <u>https://doi.org/10.1002/iroh.201011250</u>
- Descloux, S., Datry, T., Marmonier, P. 2013. Benthic and hyporheic invertebrate assemblages along a
 gradient of increasing streambed colmation by fine sediment. Aquatic Sciences, 75(4), 493-507.
 https://doi.org/10.1007/s00027-013-0295-6
- Descloux, S., Datry, T., Usseglio-Polatera, P. 2014. Trait-based structure of invertebrates along a
 gradient of sediment colmation: Benthos versus hyporheos responses. Science of the Total
 Environment, 466, 265-276. <u>https://doi.org/10.1016/j.scitotenv.2013.06.082</u>
- Di Lorenzo, T., Stoch, F., Galassi, D.M.P. 2013. Incorporating the hyporheic zone within the river
 discontinuum: longitudinal patterns of subsurface copepod assemblages in an Alpine stream.
 Limnologica, 43(4), 288-296. https://doi.org/10.1016/j.limno.2012.12.003

- Di Lorenzo, T., Fiasca, B., Di Cicco, M., Cifoni, M., Galassi, D.M.P. 2021. Taxonomic and functional trait
 variation along a gradient of ammonium contamination in the hyporheic zone of a Mediterranean
- 913 stream, *Ecological Indicators*, 132, 108268, <u>https://doi.org/10.1016/j.ecolind.2021.108268</u>.
- Dole, M. J., Chessel, D. 1986. Stabilité physique et biologique des milieux interstitiels. Cas de deux
 stations du Haut-Rhône. Annales de limnologie International Journal of Limnology, 22(1), 69-81.
- 916 https://doi.org/10.1051/limn/1986008
- Dole-Olivier, M. J., Marmonier, P. 1992. Patch distribution of interstitial communities: prevailing
 factors. Freshwater Biology, 27(2), 177-191. <u>https://doi.org/10.1111/j.1365-2427.1992.tb00532.x</u>
- 919 Dole-Olivier, M. J., Marmonier, P., Beffy, J. L. 1997. Response of invertebrates to lotic disturbance: is
- 920 the hyporheic zone a patchy refugium? Freshwater Biology, 37(2), 257-276.
 921 <u>https://doi.org/10.1046/j.1365-2427.1997.00140.x</u>
- Dole-Olivier, M. J., Galassi, D. M. P., Marmonier, P., Creuzé des Châtelliers, M. 2000. The biology and
 ecology of lotic microcrustaceans. Freshwater biology, 44(1), 63-91.
 https://doi.org/10.1046/j.1365-2427.2000.00590.x
- Dole-Olivier, M. J., Malard, F., Martin, D., Lefebure, T., Gibert, J. 2009. Relationships between
 environmental variables and groundwater biodiversity at the regional scale. Freshwater biology,
 54(4), 797-813. https://doi.org/10.1111/j.1365-2427.2009.02184.x
- Dole-Olivier, M. J. 2011. The hyporheic refuge hypothesis reconsidered: a review of hydrological
 aspects. Marine and Freshwater Research, 62(11), 1281-1302. https://doi.org/10.1071/MF11084
- 930 Dole-Olivier, M. J., Galassi, D. M., Fiers, F., Malard, F., Martin, P., Martin, D., Marmonier, P. 2015.
- Biodiversity in mountain groundwater: the Mercantour National Park (France) as a European
 hotspot. Zoosystema, *37*(4), 529-550. <u>https://doi.org/10.5252/z2015n4a1</u>
- 933 Dole-Olivier, M. J., Wawzyniak, V., Des Châtelliers, M. C., Marmonier, P. 2019. Do thermal infrared (TIR)
- 934 remote sensing and direct hyporheic measurements (DHM) similarly detect river-groundwater
- 935 exchanges? Study along a 40 km-section of the Ain River (France). Science of the Total Environment,
- 936 646, 1097-1110. <u>https://doi.org/10.1016/j.scitotenv.2018.07.294</u>
- Dudunake, T., Tonina, D., Reeder, W. J., Monsalve, A. 2020. Local and reach-scale hyporheic flow
 response from boulder-induced geomorphic changes. Water Resources Research, 56(10), 1-15.
 <u>https://doi.org/10.1029/2020WR027719</u>
- 940 Dwivedi, D., Steefel, C. I., Arora, B., Newcomer, M., Moulton, J. D., Dafflon, B., Faybishenko, B., Fox, P., 941 Nico, P., Spycher, N., Carol, R., Williams, K. H. 2018. Geochemical exports to river from the 942 intrameander hyporheic zone under transient hydrologic conditions: East River Mountainous 943 Watershed, Colorado. Water Resources Research, 54(10), 8456-8477. 944 https://doi.org/10.1029/2018WR023377

- Eme, D., Zagmajster, M., Fišer, C., Galassi, D., Marmonier, P., Stoch, F., Cornu, J.F., Oberdorf, T., Malard,
 F. 2015. Multi-causality and spatial non-stationarity in the determinants of groundwater crustacean
 diversity in Europe. Ecography, 38(5), 531-540. https://doi.org/10.1111/ecog.01092
- 948 Ercoli, F., Lefebvre, F., Delangle, M., Gode, N., Caillon, M., Raimond, R., Souty-Grosset, C. 2019. 949 Differing trophic niches of three French stygobionts and their implications for conservation of
- 950 endemic stygofauna. Aquatic Conservation: Marine and Freshwater Ecosystems, 29: 2193– 2203.
- 951 <u>https://doi.org/10.1002/aqc.3227</u>
- Fattorini, S., Fiasca, B., Di Lorenzo, T., Di Cicco, M., Galassi, D. M. P., 2020. A new protocol for assessing
 the conservation priority of groundwater-dependent ecosystems. Aquatic Conservation: Marine
 and Freshwater Ecosystems, 30: 1483–1504. <u>https://doi.org/10.1002/aqc.3411</u>
- Faulkner, B. R., Brooks, J. R., Forshay, K. J., Cline, S. P. 2012. Hyporheic flow patterns in relation to large
 river floodplain attributes. Journal of hydrology, 448, 161-173.
 https://doi.org/10.1016/j.jhydrol.2012.04.039
- Findlay, S. 1995. Importance of surface-subsurface exchange in stream ecosystems: The hyporheic
 zone. Limnology and oceanography, 40(1), 159-164. <u>https://doi-org.docelec.univ-</u>
 <u>lyon1.fr/10.4319/lo.1995.40.1.0159</u>
- 961 Franken, R. J., Storey, R. G., Williams, D. D. 2001. Biological, chemical and physical characteristics of
 962 downwelling and upwelling zones in the hyporheic zone of a north-temperate stream.
 963 Hydrobiologia, 444(1), 183-195. https://doi-org.docelec.univ-lyon1.fr/10.1023/A:1017598005228
- Gallardo, B., Gascón, S., Cabezas, Á., González-Sanchis, M., García-Antón, M., Comín, F. A. 2009.
 Relationship between invertebrate traits and lateral environmental gradients in a Mediterranean
 river-floodplain. Fundamental and Applied Limnology, 173(4), 281-292. <u>https://doi-org/</u>
 10.1127/1863-9135/2009/0173-0281
- Gayraud, S., Philippe, M. 2001. Does subsurface interstitial space influence general features and
 morphological traits of the benthic macroinvertebrate community in streams? Archiv für
 Hydrobiologie, 151(4), 667-686. https://doi-org/10.1127/archiv-hydrobiol/151/2001/667
- Gibbins, C., Vericat, D., Batalla, R. J. 2007. When is stream invertebrate drift catastrophic? The role of
 hydraulics and sediment transport in initiating drift during flood events. Freshwater Biology, 52(12),
- 973 2369-2384. <u>https://doi-org.docelec.univ-lyon1.fr/10.1111/j.1365-2427.2007.01858.x</u>
- Gomez, B. 1984. Typology of segregated (armoured/paved) surfaces: some comments. Earth Surface
 Processes and Landforms, 9(1), 19-24. <u>https://doi-org.docelec.univ-</u>
 <u>lyon1.fr/10.1002/esp.3290090103</u>
- Gomez-Velez, J. D., Wilson, J. L., Cardenas, M. B., Harvey, J. W. 2017. Flow and residence times of
 dynamic river bank storage and sinuosity-driven hyporheic exchange. Water Resources Research,
- 979 53(10), 8572-8595. <u>https://doi-org.docelec.univ-lyon1.fr/10.1002/2017WR021362</u>

Graco-Roza, C., Soininen, J., Corrêa, G., Pacheco, F. S., Miranda, M., Domingos, P., Marinho, M. M.
 2021. Functional rather than taxonomic diversity reveals changes in the phytoplankton community
 of a large dammed river. Ecological Indicators, 121, 107048.

983 <u>https://doi.org/10.1016/j.ecolind.2020.107048</u>

- Graillot, D., Paran, F., Bornette, G., Marmonier, P., Piscart, C., Cadilhac, L., 2014. Coupling groundwater
 modelling and biological indicators for identifying river/aquifer exchanges. Springerplus 3, 68.
 https://doi.org/10.1186/2193-1801-3-68
- Harbaugh, A.W., Banta, E.R., Hill, M.C., McDonald, M.G., 2000. Modflow-2000, the U.S. Geological
 survey modular Ground-Water model user guide to modularization concepts and the groundwater flow process. U.S. Geological Survey, Open-File Report 00-92, 1-121, 26p.
- Harvey, J. W., Böhlke, J. K., Voytek, M. A., Scott, D., Tobias, C. R. 2013. Hyporheic zone denitrification:
 Controls on effective reaction depth and contribution to whole-stream mass balance. Water
- 992 Resources Research, *49*(10), 6298-6316. <u>https://doi-org.docelec.univ-lyon1.fr/10.1002/wrcr.20492</u>
- 993 Hunt, W. G., Stanley, E. H. 2003. Environmental factors influencing the composition and distribution of
- the hyporheic fauna in Oklahoma streams: Variation across ecoregions. Archiv fur Hydrobiologie,
 158(1), 1-23. <u>https://doi-org/10.1127/0003-9136/2003/0158-0001</u>
- 996 Hutchins, B. T., Swink, A. P., Diaz, P. H., Schwartz, B. F. 2020. Environmental influences on invertebrate
- 997 diversity and community composition in the hyporheic zone ecotone in Texas, USA: contrasts
- between co-occurring epigean taxa and stygobionts. Hydrobiologia, 847(19), 3967-3982.
 <u>https://doi-org.docelec.univ-lyon1.fr/10.1007/s10750-020-04379-8</u>
- Hynes, H. B. N. 1975. The stream and its valley. Internationale Vereinigung für theoretische und
 angewandte Limnologie: Verhandlungen, 19(1), 1-15. <u>https://doi-org.docelec.univ-</u>
 lyon1.fr/10.1080/03680770.1974.11896033
- Jain, S. C. 1990. Armor or pavement. Journal of Hydraulic Engineering, 116(3), 436-440.
 <u>https://doi.org/10.1061/(ASCE)0733-9429(1990)116:3(436)</u>
- Jones Jr, J. B., Fisher, S. G., Grimm, N. B. 1995. Nitrification in the hyporheic zone of a desert stream
 ecosystem. Journal of the North American Benthological Society, 14(2), 249-258.
 <u>https://doi.org/10.2307/1467777</u>
- Juget, J., Lafont, M. 1994. Theoretical habitat templets, species traits, and species richness: aquatic
 oligochaetes in the Upper Rhône River and its floodplain. Freshwater Biology, 31(3), 327-340.
 <u>https://doi-org.docelec.univ-lyon1.fr/10.1111/j.1365-2427.1994.tb01744.x</u>
- Lawton, J. H. 1994. What do species do in ecosystems? Oikos, 71(3), 367-374. <u>https://doi-</u>
 org.docelec.univ-lyon1.fr/10.2307/3545824

- Lefebvre, S., Marmonier, P., Pinay, G., Bour, O., Aquilina, L., Baudry, J. 2005. Nutrient dynamics in
 interstitial habitats of low-order rural streams with different bedrock geology. Archiv fur
 Hydrobiologie, 164(2), 169-191. https://doi.org/10.1127/0003-9136/2005/0164-0169
- Leigh, C., Bonada, N., Boulton, A. J., Hugueny, B., Larned, S. T., Vander Vorste, R., Datry, T. 2016.
 Invertebrate assemblage responses and the dual roles of resistance and resilience to drying in
 intermittent rivers. Aquatic Sciences, 78(2), 291-301. <u>https://doi-org.docelec.univ-</u>
 lyon1.fr/10.1007/s00027-015-0427-2
- Lejot, J. 2008. Suivi des formes fluviales par télédétection à très haute résolution. Application aux
 programmes de restauration de la basse vallée de l'Ain et du haut-Rhône (Chautagne). PhD thesis,
 University of Lyon 2, 257p.
- Lewandowski, J., Arnon, S., Banks, E., Batelaan, O., Betterle, A., Broecker, T., et al. 2019. Is the
 hyporheic zone relevant beyond the scientific community? Water, 11(11), 2230. <u>https://doi-org</u>
 /0.3390/w1112230
- Lin, Q., Song, J., Gualtieri, C., Cheng, D., Su, P., Wang, X., Fu, J., Peng, J. 2020. Effect of Hyporheic
 Exchange on Macroinvertebrate Community in the Weihe River Basin, China. Water, 12(2), 457.
 https://doi-org/10.3390/w12020457
- Magliozzi, C., Grabowski, R. C., Packman, A. I., Krause, S. 2018. Toward a conceptual framework of
 hyporheic exchange across spatial scales. Hydrology and Earth System Sciences, 22(12), 6163-6185.
 https://doi.org/10.5194/hess-22-6163-2018
- Magliozzi, C, Usseglio-Polatera, P, Meyer, A, Grabowski, RC. 2019. Functional traits of hyporheic and
 benthic invertebrates reveal importance of wood-driven geomorphological processes in rivers.
 Functional Ecology, 33: 1758– 1770. <u>https://doi.org/10.1111/1365-2435.13381</u>
- 1035 Malard, F., Galassi, D., Lafont, M., Doledec, S., Ward, J. V. 2003. Longitudinal patterns of invertebrates
- in the hyporheic zone of a glacial river. Freshwater Biology, 48(10), 1709-1725. <u>https://doi-org.docelec.univ-lyon1.fr/10.1046/j.1365-2427.2003.01118.x</u>
- Malcolm, I. A., Soulsby, C., Youngson, A. F., Hannah, D. M. 2005. Catchment-scale controls on groundwater–surface water interactions in the hyporheic zone: implications for salmon embryo survival. River Research and Applications, 21(9), 977-989. <u>https://doi-org.docelec.univ-</u>
 lyon1.fr/10.1002/rra.861
- Mao, L., Surian, N. 2010. Observations on sediment mobility in a large gravel-bed river.
 Geomorphology, 114(3), 326-337. <u>https://doi.org/10.1016/j.geomorph.2009.07.015</u>
- Marion, A., Packman, A. I., Zaramella, M., Bottacin-Busolin, A. 2008. Hyporheic flows in stratified beds.
 Water Resources Research, 44(9), 1-10. <u>https://doi-org.docelec.univ-</u>
 lyon1.fr/10.1029/2007WR006079

- Marmonier, P., Vervier, P., Giber, J., Dole-Olivier, M. J. 1993. Biodiversity in ground waters. Trends in
 Ecology & Evolution, 8(11), 392-395. <u>https://doi.org/10.1016/0169-5347(93)90039-R</u>
- 1049Marmonier, P., Bodergat, A. M., Doledec, S. 1994. Theoretical habitat templets, species traits, and1050species richness: ostracods (Crustacea) in the Upper Rhône River and its floodplain. Freshwater
- 1051
 biology,
 31(3),
 341-355.
 https://doi-org.docelec.univ-lyon1.fr/10.1111/j.1365

 1052
 2427.1994.tb01745.x
- Marmonier, P., Piscart, C., Sarriquet, P. E., Azam, D., Chauvet, E. 2010a. Relevance of large litter bag
 burial for the study of leaf breakdown in the hyporheic zone. Hydrobiologia, 641(1), 203-214.
 https://doi-org.docelec.univ-lyon1.fr/10.1007/s10750-009-0082-5
- 1056 Marmonier, P., Luczyszyn, H., Des Chatelliers, M. C., Landon, N., Claret, C., Dole-Olivier, M. J. 2010b.
- 1057 Hyporheic flowpaths and interstitial invertebrates associated with stable and eroded river sections:
- interactions between micro-and meso-scales. Fundamental and Applied Limnology, 176(4), 303317. https://doi.org/10.1127/1863-9135/2010/0176-0303
- Marmonier, P., Archambaud, G., Belaidi, N., Bougon, N., Breil, P., Chauvet, E., Claret, C., Cornut, J.,
 Datry, T. , Dole-Olivier, M.-J., Dumont, B., Flipo, N., Foulquier, A., Gérino, M., Guilpart, A., Julien, F.,
 Maazouzi, C., Martin, D. , Mermillod-Blondin, F., Montuelle, B., Namour, Ph., Navel, S., Ombredane,
- 1063 D., Pelte, T., Piscart, C., Pusch, M., Stroffek, S., Robertson, A., Sanchez-Pérez, J.-M., Sauvage, S.,
- Taleb, A., Wantzen, M., Vervier, P. 2012. The role of organisms in hyporheic processes: gaps in
 current knowledge, needs for future research and applications. Annales de LimnologieInternational Journal of Limnology, 48(3), 253-266. <u>https://doi.org/10.1051/limn/2012009</u>
- Marmonier, P., Olivier, M.J., Creuzé des Châtelliers, M., Paran, F., Graillot, D., Winiarski, T., Konecny Dupré, L., Navel, S., Cadilhac, L., 2019. Does spatial heterogeneity of hyporheic fauna vary similarly
 with natural and artificial changes in braided river width? Science of the Total Environment, 689,
 57–69. https://doi.org/10.1016/j.scitotenv.2019.06.352
- Marmonier, P., des Châtelliers, M. C., Dole-Olivier, M. J., Radakovitch, O., Mayer, A., Chapuis, H.,
 Graillot, D., Re-Bahuaud, J., Johannet, A., Cadilhac, L. 2020. Are surface water characteristics
 efficient to locate hyporheic biodiversity hotspots? Science of the Total Environment, 738, 139930.
 https://doi.org/10.1016/j.scitotenv.2020.139930
- Marzadri, A., Tonina, D., Bellin, A. 2012. Morphodynamic controls on redox conditions and on nitrogen
 dynamics within the hyporheic zone: Application to gravel bed rivers with alternate-bar
 morphology. Journal of Geophysical Research: Biogeosciences, 117(G3), 1-14. https://doi-org.docelec.univ-lyon1.fr/10.1029/2012JG001966
- Mathers, K. L., Hill, M. J., Wood, P. J. 2017. Benthic and hyporheic macroinvertebrate distribution
 within the heads and tails of riffles during baseflow conditions. Hydrobiologia, 794(1), 17-30.
 https://doi-org.docelec.univ-lyon1.fr/10.1007/s10750-017-3092-8

- Mermillod-Blondin, F., Creuzé des Châtelliers, M., Gerino, M., Gaudet, J. P. 2000. Testing the effect of
 Limnodrilus sp. (Oligochaeta, Tubificidae) on organic matter and nutrient processing in the
 hyporheic zone: a microcosm method. Archiv für Hydrobiologie, 149(3), 467-487.
 <u>https://doi.org/10.1127/archiv-hydrobiol/149/2000/467</u>
- Mermillod-Blondin, F., Rosenberg, R. 2006. Ecosystem engineering: the impact of bioturbation on
 biogeochemical processes in marine and freshwater benthic habitats. Aquatic sciences, 68(4), 434 442. https://doi-org.docelec.univ-lyon1.fr/10.1007/s00027-006-0858-x
- Mermillod-Blondin, F., Winiarski, T., Foulquier, A., Perrissin, A., Marmonier, P. 2015. Links between
 sediment structures and ecological processes in the hyporheic zone: ground-penetrating radar as a
 non-invasive tool to detect subsurface biologically active zones. Ecohydrology, 8(4), 626-641.
 https://doi-org.docelec.univ-lyon1.fr/10.1002/eco.1530
- Michel, G., Malard, F., Deharveng, L., Di Lorenzo, T., Sket, B., De Broyer, C. 2009. Reserve selection for
 conserving groundwater biodiversity. Freshwater Biology, 54(4), 861-876. <u>https://doi-</u>
 org.docelec.univ-lyon1.fr/10.1111/j.1365-2427.2009.02192.x
- Navel, S., Mermillod-Blondin, F., Montuelle, B., Chauvet, E., Simon, L., Piscart, C., Marmonier, P. 2010.
 Interactions between fauna and sediment control the breakdown of plant matter in river
 sediments. Freshwater Biology, 55(4), 753-766. <u>https://doi-org.docelec.univ-</u>
 lyon1.fr/10.1111/j.1365-2427.2009.02315.x
- Navel, S., Mermillod-Blondin, F., Montuelle, B., Chauvet, E., Simon, L., Marmonier, P. 2011. Water–
 sediment exchanges control microbial processes associated with leaf litter degradation in the
 hyporheic zone: a microcosm study. Microbial ecology, 61(4), 968-979. https://doi-org.docelec.univ-lyon1.fr/10.1007/s00248-010-9774-7
- Nelson, A. R., Sawyer, A. H., Gabor, R. S., Saup, C. M., Bryant, S. R., Harris, K. D., Briggs M.A., Williams
 K.H., Wilkins, M. J. 2019. Heterogeneity in hyporheic flow, pore water chemistry, and microbial
- 1106 community composition in an alpine streambed. Journal of Geophysical Research: Biogeosciences,
- 1107 124(11), 3465-3478. <u>https://doi-org.docelec.univ-lyon1.fr/10.1029/2019JG005226</u>
- Nogaro, G., Datry, T., Mermillod-Blondin, F., Descloux, S., Montuelle, B. 2010. Influence of streambed
 sediment clogging on microbial processes in the hyporheic zone. Freshwater biology, 55(6), 1288 1302. https://doi-org.docelec.univ-lyon1.fr/10.1111/j.1365-2427.2009.02352.x
- 1111 Nogaro, G., Burgin, A. J. 2014. Influence of bioturbation on denitrification and dissimilatory nitrate
- reduction to ammonium (DNRA) in freshwater sediments. Biogeochemistry, 120(1), 279-294.
 <u>https://doi-org.docelec.univ-lyon1.fr/10.1007/s10533-014-9995-9</u>
- Olsen, D. A., Townsend, C. R. 2003. Hyporheic community composition in a gravel-bed stream:
 influence of vertical hydrological exchange, sediment structure and physicochemistry. Freshwater

- 1116
 Biology,
 48(8),
 1363-1378.
 <u>https://doi-org.docelec.univ-lyon1.fr/10.1046/j.1365-</u>

 1117
 2427.2003.01097.x
- Orghidan, T. (1959). Ein neuer Lebensraum des unterirdischen Wassers: der hyporheische Biotop.
 Archiv für Hydrobiologie, 55(3), 392-414.
- O'Sullivan, M., Huallachain, D. O., Antunes, P. O., Jennings, E., Kelly-Quinn, M. 2019. The impacts of
 cattle access to headwater streams on hyporheic zones. Biology and Environment: Proceedings of
 the Royal Irish Academy, 119(1), 13-27. https://doi.org/10.3348/BIOE.2019.02
- 1123 Paillex, A., Dolédec, S., Castella, E., Mérigoux, S., Aldridge, D. C. 2013. Functional diversity in a large
- river floodplain: anticipating the response of native and alien macroinvertebrates to the restoration
 of hydrological connectivity. Journal of Applied Ecology, 50(1), 97-106. <u>https://doi-</u>
 org.docelec.univ-lyon1.fr/10.1111/1365-2664.12018
- Palmer, M. A., Bely, A. E., Berg, K. E. 1992. Response of invertebrates to lotic disturbance: a test of the
 hyporheic refuge hypothesis. Oecologia, 89(2), 182-194. <u>https://doi-org.docelec.univ-</u>
 lyon1.fr/10.1007/BF00317217
- Paran, F., Augeard, B., 2017. Guide technique Interactions nappe/rivière: des outils pour comprendre
 et mesurer les échanges. Guides et protocoles. Agence française pour la biodiversité, pp. 1–102.
- Parker, J., Cao, Y., Sass, G. G., Epifanio, J. 2018. Large river fish functional diversity responses to
 improved water quality over a 28 year period. Ecological indicators, 88, 322-331.
 <u>https://doi.org/10.1016/j.ecolind.2018.01.035</u>
- Peck, D. V., Lazorchak, J. M. Klemm, D. J. 2000. Environmental monitoring and assessment programsurface water: Western pilot study field operations manual for wadeable streams. US
 Environmental protection Agency, Washington D.C.
- 1138 Peralta-Maraver, I., Reiss, J., Robertson, A. L. 2018. Interplay of hydrology, community ecology and
- pollutant attenuation in the hyporheic zone. Science of the Total Environment, 610-611, 267-275.
 <u>https://doi.org/10.1016/j.scitotenv.2017.08.036</u>
- 1141Peterson, E. W., Sickbert, T. B., Moore, S. L. 2008. High frequency stream bed mobility of a low-gradient1142agricultural stream with implications on the hyporheic zone. Hydrological Processes, 22(21), 4239-
- 1143 4248. <u>https://doi-org.docelec.univ-lyon1.fr/10.1002/hyp.7031</u>
- Petts, G. E., Amoros, C. 1996. The fluvial hydrosystem, Springer, Dordrecht. <u>https://doi-</u>
 <u>org.docelec.univ-lyon1.fr/10.1007/978-94-009-1491-9 1</u>
- 1146 Poff LeRoy, N. L. 1997. Landscape filters and species traits: towards mechanistic understanding and
- 1147 prediction in stream ecology. Journal of the North American Benthological Society, 16(2), 391-409.
- 1148 https://doi.org/10.2307/1468026

- Poole, G. C. 2002. Fluvial landscape ecology: addressing uniqueness within the river discontinuum.
 Freshwater Biology, 47(4), 641-660. <u>ttps://doi-org.docelec.univ-lyon1.fr/10.1046/j.1365-</u>
 <u>2427.2002.00922.x</u>
- Poole, G. C. 2010. Stream hydrogeomorphology as a physical science basis for advances in stream
 ecology. Journal of the North American Benthological Society, 29(1), 12-25.
 <u>https://doi.org/10.1899/08-070.1</u>
- 1155 R Core Team, 2021. R: A language and environment for statistical computing. R Foundation for
 1156 Statistical Computing, Vienna, Austria. URL <u>https://www.R-project.org/.</u>
- Richards, C., Bacon, K. L. 1994. Influence of fine sediment on macroinvertebrate colonization of surface
 and hyporheic stream substrates. The Great Basin Naturalist, 54(2), 106-113.
 http://www.jstor.org/stable/41712819
- Rollet, A. J., Piégay, H., Dufour, S., Bornette, G., Persat, H. 2014. Assessment of consequences of
 sediment deficit on a gravel river bed downstream of dams in restoration perspectives: application
- of a multicriteria, hierarchical and spatially explicit diagnosis. River Research and Applications,
 30(8), 939-953. <u>https://doi-org.docelec.univ-lyon1.fr/10.1002/rra.2689</u>
- Roy Chowdhury, S., Zarnetske, J. P., Phanikumar, M. S., Briggs, M. A., Day-Lewis, F. D., Singha, K. 2020.
 Formation criteria for hyporheic anoxic microzones: Assessing interactions of hydraulics, nutrients,
 and biofilms. Water Resources Research, 56(3), 1-15. <u>https://doi-org.docelec.univ-</u>
 lyon1.fr/10.1029/2019WR025971
- Schmera, D., Heino, J., Podani, J., Erős, T., Dolédec, S. 2017. Functional diversity: a review of
 methodology and current knowledge in freshwater macroinvertebrate research. Hydrobiologia,
 787(1), 27-44. https://doi-org.docelec.univ-lyon1.fr/10.1007/s10750-016-2974-5
- Schmid-Araya, J. M. 2000. Invertebrate recolonization patterns in the hyporheic zone of a gravel
 stream. Limnology and Oceanography, 45(4), 1000-1005. <u>https://doi-org.docelec.univ-</u>
 <u>lyon1.fr/10.4319/lo.2000.45.4.1000</u>
- Sennatt, K. M., Salant, N. L., Renshaw, C. E., Magilligan, F. J. 2006. Assessment of Methods for
 Measuring Embeddedness: Application to Sedimentation in Flow Regulated STREAMS1. Journal of
 the American Water Resources Association, 42(6), 1671-1682. <u>https://doi-org.docelec.univ-</u>
- 1177 lyon1.fr/10.1111/j.1752-1688.2006.tb06028.x
- Shrivastava, S., Stewardson, M. J., Arora, M. 2021a. Influence of bioturbation on hyporheic exchange
 in streams: Conceptual model and insights from laboratory experiments. Water Resources
 Research, 57, e2020WR028468. <u>https://doi.org/10.1029/2020WR028468</u>
- Shrivastava, S., Stewardson, M. J., Arora, M. 2021b. Sediment reworking in streambeds with fine
 sediment deposits and its influence on hyporheic flow regime. Water Resources Research, 57,
 e2021WR030360. https://doi.org/10.1029/2021WR030360

- Singer, M. B. 2008. Downstream patterns of bed material grain size in a large, lowland alluvial river
 subject to low sediment supply. Water Resources Research, 44(12). <u>https://doi-org.docelec.univ-</u>
 lyon1.fr/10.1029/2008WR007183
- 1187 Słowik, M., Kiss, K., Czigány, S., Gradwohl-Valkay, A., Dezso, J., Halmai, A., Marciniak, A., Tritt R.,
- Pirkhoffer, E. 2021. The influence of changes in flow regime caused by dam closure on channel
 planform evolution: insights from flume experiments. Environmental Earth Sciences, 80, 165.
 https://doi.org/10.1007/s12665-021-09437-5
- Snelder, T. H., Lamouroux, N., Pella, H. 2011. Empirical modelling of large scale patterns in river bed
 surface grain size. Geomorphology, 127(3-4), 189-197.
 https://doi.org/10.1016/j.geomorph.2010.12.015
- 1194 Stonedahl, S. H., Harvey, J. W., Packman, A. I. 2013. Interactions between hyporheic flow produced by
- stream meanders, bars, and dunes. Water Resources Research, 49(9), 5450-5461. <u>https://doi-org.docelec.univ-lyon1.fr/10.1002/wrcr.20400</u>
- Storey, R. G., Williams, D. D., Fulthorpe, R. R. 2004. Nitrogen processing in the hyporheic zone of a
 pastoral stream. Biogeochemistry, 69(3), 285-313. <u>https://doi-org.docelec.univ-</u>
 lyon1.fr/10.1023/B:BIOG.0000031049.95805.ec
- Strayer, D. L., May, S. E., Nielsen, P., Wollheim, W., Hausam, S. 1997. Oxygen, organic matter, and
 sediment granulometry as controls on hyporheic animal communities. Archiv für Hydrobiologie,
 140(1), 131-144.
- Stubbington, R., Dole-Olivier, M. J., Galassi, D. M., Hogan, J. P., Wood, P. J. 2016. Characterization of
 macroinvertebrate communities in the hyporheic zone of river ecosystems reflects the pump sampling technique used. PloS one, 11(10), e0164372.
 https://doi.org/10.1371/journal.pone.0164372
- Tachet, H., Richoux, P. H., Bournaud, M., Usseglio-Polatera, P. 2000. Invertébrés d'eau douce.
 Systématique, biologie, écologie. CNRS éditions, Paris.
- Tilman, D., Knops, J., Wedin, D., Reich, P., Ritchie, M., Siemann, E. 1997. The influence of functional
 diversity and composition on ecosystem processes. Science, 277(5330), 1300-1302.
 <u>https://doi.org/10.1126/science.277.5330.1300</u>
- 1212 Tonina, D., Buffington, J. M. 2009. Hyporheic exchange in mountain rivers I: Mechanics and 1213 environmental effects. Geography Compass, 3(3), 1063-1086. <u>https://doi-org.docelec.univ-</u> 1214 lyon1.fr/10.1111/j.1749-8198.2009.00226.x
- 1215 Tonina, D., de Barros, F. P., Marzadri, A., Bellin, A. 2016. Does streambed heterogeneity matter for
- hyporheic residence time distribution in sand-bedded streams? Advances in Water Resources, 96,
 120-126. https://doi.org/10.1016/j.advwatres.2016.07.009

- 1218 Triska, F. J., Duff, J. H., Avanzino, R. J. 1993. The role of water exchange between a stream channel and
- its hyporheic zone in nitrogen cycling at the terrestrial—aquatic interface, in Hillbricht-Ilkowska, A.,
- 1220 Pieczynska, E. (Eds), Nutrient dynamics and retention in land/water ecotones of lowland, temperate
- 1221
 lakes and rivers.
 Springer,
 Dordrecht,
 pp.
 167-184.
 https://doi-org.docelec.univ

 1222
 lyon1.fr/10.1007/978-94-011-1602-2_20
- 1223 Usseglio-Polatera, P., Bournaud, M., Richoux, P., Tachet, H. 2000. Biological and ecological traits of

benthic freshwater macroinvertebrates: relationships and definition of groups with similar traits.

- 1225 Freshwater Biology, 43(2), 175-205. <u>https://doi-org.docelec.univ-lyon1.fr/10.1046/j.1365-</u> 1226 2427.2000.00535.x
- Vadher, A. N., Stubbington, R., Wood, P. J. 2015. Fine sediment reduces vertical migrations of
 Gammarus pulex (Crustacea: Amphipoda) in response to surface water loss. Hydrobiologia, 753(1),
 61-71. <u>https://doi-org.docelec.univ-lyon1.fr/10.1007/s10750-015-2193-5</u>
- Valett, H. M., Hakenkamp, C. C., Boulton, A. J. 1993. Perspectives on the hyporheic zone: integrating
 hydrology and biology. Introduction. Journal of the North American Benthological Society, 12(1),
 40-43. <u>https://doi.org/10.2307/1467683</u>
- Vander Vorste, R. V., Corti, R., Sagouis, A., Datry, T. 2016. Invertebrate communities in gravel-bed,
 braided rivers are highly resilient to flow intermittence. Freshwater Science, 35(1), 164-177.
 <u>https://doi.org/10.1086/683274</u>
- Williams, D. D. 1984. The hyporheic zone as an habitat for aquatic insects and associated arthropods,
 in Resh, V.H., Rosenberg, D.M. (Eds), The ecology of aquatic insects, Praeger Scientific, New York,
 pp.430-455.
- Williams, D. D. 1993. Nutrient and flow vector dynamics at the hyporheic/groundwater interface and
 their effects on the interstitial fauna, in Hillbricht-Ilkowska, A., Pieczynska, E. (Eds), Nutrient
 dynamics and retention in land/water ecotones of Lowland, Temperate Lakes and Rivers, Springer,
- 1242 Dordrecht, pp. 185-198. <u>https://doi.org/10.1007/978-94-011-1602-2_21</u>
- Williams, D. D., Febria, C. M., Wong, J. C. 2010. Ecotonal and other properties of the hyporheic zone.
 Fundamental and Applied Limnology, 176(4), 349-364. <u>https://doi.org/10.1127/1863-</u>
 9135/2010/0176-0349
- Wood, P. J., Armitage, P. D. 1997. Biological effects of fine sediment in the lotic environment.
 Environmental management, 21(2), 203-217. <u>https://doi.org/10.1007/s002679900019</u>
- Zheng, L., Cardenas, M. B., Wang, L., Mohrig, D. 2019. Ripple effects: Bed form morphodynamics
 cascading into hyporheic zone biogeochemistry. Water Resources Research, 55(8), 7320-7342.
 https://doi-org.docelec.univ-lyon1.fr/10.1029/2018WR023517
- 1251

1224

Table 1: Comparison of the effects of local hydrology, sediment grain size and sediment stability on the functional traits of the hyporheic assemblages. One way ANOVAs were performed with stations as replicates (mean values for all depths and replicates, after log transformation; n.s. above the significant level, - no test possible for food type groups because a species may belong to more than one trophic group, see histogram in Supplementary material Fig. S4).

1261

1255

	Local hydrology ⁽¹⁾				Grain size ⁽²⁾		Stabilit	y ⁽³⁾
	Abundanc	es	Richness		Abd	Rich	Abd	Rich
	F _(2,27)	р	F _(2,27)	р	F _(2,27)	F _(2,27)	F _(1,28)	F _(1,28)
Total fauna	4.97	0.014	4.96	0.014	n.s.	n.s.	n.s.	n.s.
Ecological groups								
Temp. hyporheos	12.05	0.0002	8.94	0.001	n.s.	n.s.	n.s.	n.s.
Perm. hyporheos		n.s.	3.51	0.04	n.s.	n.s.	n.s.	n.s.
Stygobites		n.s.	6.83	0.004	n.s.	n.s.	n.s.	n.s.
Food type groups								
Sediment feeders	5.40	0.011	-	-	n.s.	-	n.s.	-
POM feeders	4.69	0.017	-	-	n.s.	-	n.s.	-
Algae feeders	3.47	0.04	-	-	n.s.	-	n.s.	-
Preadators		n.s.	-	-	n.s.	-	n.s.	-

1262

⁽¹⁾ Local hydrology are downwelling (n=16), low exchange (n=4) and upwelling contexts (n=10).

⁽²⁾ Sediment grain size classes are fine (n=13), mixed (n=10) and coarse surface sediment (n=7).

⁽³⁾ Sediment stability are with (n=17) or without biofilm development (n=13).

1266

1268 Figure legends:

Figure 1: Study area (A), the Lower Ain River from Pont d'Ain to the confluence with the Rhône River (background elevation map from Geographic National Institute, Géoportail). Blue lines: Ain and Rhône river channels; pointed dark line: upstream eroded sector before 2005; dotted lines: limits of the morainic hills. Location of the 15 sampled gravel bars (A), upstream and downstream stations at each gravel bar (B) and sampling design at each station (C).

1274

Figure 2. Chemical characteristics of hyporheic water, sediment grain size and sediment stability. Examples showing the classification of the stations into the three groups of stream-groundwater exchanges according to electric conductivity and temperature of the surface and hyporheic waters: A-Upwellings (UPW), B- Low exchanges (LOW EX), C- Downwelling (DNW). Examples of sediment characteristics: D- fine sediment, E- mixed, F- coarse. Example of biofilm development on unstable (G) or stable (H) benthic layer. List of the stations in each group with U for upstream and D for downstream positions along the gravel bars (see Fig. 1).

1282

Figure 3: Hydrogeological characteristics of the studied sector based on geomatic analysis of the study reach (A) and details of the eight stations with upwelling zones as derived by the hyporheic water chemistry (B).

1286

Figure 4. Principal Component Analysis on hyporheic water characteristics. Correlation circle of physico-chemical parameters (A) Cond-Electric conductivity, VGH-Vertical Hydraulic Gradient, O₂-Dissolved oxygen, T°C-Temperature. Location of the sampling points on the Axis 1 x Axis 2 biplots, according to the local hydrology (B), the grain size (C) and the sediment stability (D). Oxygen concentrations by groups and depths (means and standard errors) for local hydrology groups (E), grain size groups (F) and stability groups (G) with results of ANOVA tests between groups.

1293

Figure 5: Longitudinal distribution of hyporheic assemblages. Means (and standard errors) for the 15 gravel bars and positions (i.e. 30 stations) for total abundance and taxonomic richness (left panels), for ecological groups based on affinities to groundwater (center panels) and for trophic functional groups (right panels).

Figure 6. Correspondance analysis on hyporheic assemblages (the 1st axis represented 6% and the 2nd axis 5.3% of the total inertia). Location of the taxa in the 1st and 2nd axes is separated in three major taxonomic groups: (A) oligochaetes, molluscs, leeches, (B) crustaceans and (C) insects, with stygobite species in blue and other hyporheic species in black. The location of all samples between the 1st and 1302 2nd axes also separated by hydrological exchange groups (D), sediment grain size groups (E) and
 1303 sediment stability groups (F).

- Figure 7: Rarefaction curves of the taxonomic richness in different hydrological contexts (Dwn:
 downwelling zones, Upw: upwelling zones and LowEX: low exchange zones). Shaded areas represented
 confidence intervals.
- 1307

Figure 8: Taxonomic richness (left panels), abundances of the total fauna and of the 3 functional groups based on affinities with groundwater (center panels), and abundances of the food type groups (right panels) in different hydrological contexts and depths (means and standard errors). Differences between hydrological contexts (H) and depths (D) tested by a two-ways ANOVA with *<5%, **<1%, ***<0.1%, n.s. non significant.

1313

Figure 9. Functional diversity for ecological groups (A) and food type groups (B) for the longitudinal trend (left) and the combination of depth and hydrological contexts (right, means and standard errors). The ecological group diversity significantly correlated with the distance (in km) with a polynomial regression curve ($y = 0.0021 x^2 + 0.075 x + 1.3$), while the food type group diversity significantly fitted with a linear regression curve (y = 0.023 x + 2.396). Differences between hydrological contexts (H) and depths (D) tested by a two-ways ANOVA with *<5%, **<1%, ***<0.1%, n.s. not significant.

1320

























1366 Supplementary material: Tables.

1367 Table S1. Codes for the taxa used in the Factorial Analysis (Stygobite species in red).

Taxa namesCodesTaxa namesCodesAedosonstidaeAelosValvato sp.Valvato sp.Valvato sp.Valvato sp.Valvato sp.Valvato sp.Valvato sp.Valvato sp.Sylvatili <th></th> <th></th> <th></th> <th></th> <th></th> <th></th>						
Aeolosomatidae Valvata sp. Valvata Cyndropsix value C.vidua Lumbrickae Lumbrick Bitymine Potromory is variegata P.variegata Darydnius micheelseni Dorydni Gymulus sp. Gyraul Gammorus sp. (javeniles) G.pulli Stylodnilus sp. Titcho Bythiospecuni d diaphanum Bythiospecuni S (javeniles) Gammorus sp. (javeniles) Niphargus Chactopster diaphanus C.diaph Stylum sp. Pisidum sp. Pisidum sp. Niphargus sp. (javeniles) Nishargus Nisha	Taxa names	Codes	Taxa names	Codes	Taxa names	Codes
Lumbric Dorydriu SynchesBithynia tentsculataBithynia tentsculataPotamocynis voriegal Sonulia SynchesAcra and Sonulia Synches <th< td=""><td>Aeolosomatidae</td><td>Aelos</td><td>Valvata sp.</td><td>Valvata</td><td>Cypridopsis vidua</td><td>C.vidua</td></th<>	Aeolosomatidae	Aelos	Valvata sp.	Valvata	Cypridopsis vidua	C.vidua
Dorydnik michaelseni Dorydnik Gyraulu Gyraulu Garmarus pulex G. gulex Styladrikus sp. Trichod III Bythiospeund diaphonus Garmarus pulex G. gulex Cheetogaster diaphonus C. diaph Strichol III Mythargus sp. (inventiles) Nishargus Cheetogaster diastrophus C. diaph Strichik d'Utree Spiridium Niphargus sp. (inventiles) Nickorhoux Nois barbato N. harb Aranthocyclops resustus A venustus Nighargus cs.popril Nickorhoux Nois barbato N. harb Aranthocyclops resustus A venustus Nighargus cs.popril Nickorhoux Nois barbato N.barb Aranthocyclops resustus A venustus Nighargus cs.popril Nickorhoux Nais pardiais N.pard Macrocyclops abidus Microchorao Microchorao Sylaria locustris Sylaria Eucyclops macruroides Emacrur Parabethyreello c1 stygia Bathyreel Pristina equipatie Fristage parabethyreello c1 stygia Bathyreel Liperclops parabethyreello c1 stygia Bathyreel Nighargus foreli	Lumbricidae	Lumbric	Bithynia tentaculata	Bithyn	Potamocypris variegata	P.variegata
Stylod Tricho Bytkospeur d'Iduphonus Islamia Cammarus puixes G. pulex Choctopaster diaphonus C. Glasst Paidium sp. Spiralik Niphargus Sp. (unenles) Ni chorto Nois brothot Nibar Acanthocyclops rebustus Aroustus Niphargus gr. fonctus Ni chorto Nois brothot Nibar Acanthocyclops rebustus Aroustus Niphargus gr. fonctus Nicon Nois communis N.com Acanthocyclops rebustus Aroustus Niphargus gr. fonctus Nicon Nois drogis N.pard Megocyclops viridis M.Niridis Probathymeliot Stypia Bathiteri Nois drogis N.pard Megocyclops viridis M.iniridis Acentaria Probathymeliot Stypia Bathiteri Stylaria lacustris Styla Excyclops sterulatus Exercul Placetaria Probathymeliot Stypia Bathice Stypia Stylaria lacustris Diacyclops putenlis Diacyclops putenlis Diacyclops put	Dorydrilus michaelseni	Dorydril	Gyraulus sp.	Gyraul	Gammarus sp. (juveniles)	G.pulli
Trichodius sp.TrichoBythiospum di diaphonumBythiospGammarus fossarumG. fossarChectogaster diaphonusC. CalaphSpiridiuk d' VitreeSpiridiukNiphargus sp. (uveniles)Niphargus sp. (uveniles)Niphargus sp. (uveniles)Niphargus sp. (uveniles)Ni hcohNais barbataN. NarbAcanthocyclops resustusA venustusNiphargus sp. (uveniles)Ni AcasparNois barbataN. NarbA canthocyclops resustusA venustusNiphargus coparyiN. CasparNois barbataN. NarbA canthocyclops resistivusA senustusNiphargus coparyiN. CasparNois barbataN. NarbaA canthocyclops resistivusA senitivus sp.A selius sp.A selius sp.Nois pordolisN. PardMacrocyclops resistivusA senitivus sp.A selius sp.A seliusNois pordolisN. PardMacrocyclops retrulatusE serrulParabathynelio cl styginBathynelStylario lacustrisStylaEucyclops parcurolesE marcurPecoparea [uveniles]PiecoStylario lacustrisStylaEucyclops parcurolesE marcurPecoparea [uveniles]PiecoStylario lacustrisStylaEucyclops parcurolesE marcurPecoparea [uveniles]PiecoStylario lacustrisStylaEucyclops parcurolesE marcurPecoparea [uveniles]PiecoStylario lacustrisStylaEucyclops parcurolesE marcurPiecoEucyclopsStylario lacustrisStylarioDupyclops belgicusDubgicus	Stylodrilus sp.	Stylo	Islamia cf minuta	Islamia	Gammarus pulex	G.pulex
Chectopster diapharus C. Glajat Spiratik Tvitren Spiratik Niphargus chenorhadonenis Niphargus chenorhadonenis Ni hanon Nais alpino Ni alpi Hydracanina Hydrac Niphargus gr. lockbannenis Ni hanon Nais bretscheri N. bret Acanthocyclops venustus A venustus Miphargus gr. lockbannenis Ni Acanthocyclops sobustus A venustus Niphargus gr. lockbannenis Ni Acasta Nais di upin Niphargus gr. lockbannenis	Trichodrilus sp.	Tricho	Bythiospeum cf diaphanum	Bythiosp	Gammarus fossarum	G.fossar
Chaetogaster disstrophus C. diast Pisdium sp. Pisdium Nphargus transhondanensis N-theno Nais alpina N. Natp Acanthocyclogs venusus A venusus Nphargus gr.kochionus N.koch Nais barbata N. Natp A canthocyclogs venusus A venusus Nphargus gr.honthonus N.koch Nais prestoreri N. Nort A canthocyclogs sensitivus A sensitivus Salentinella sp. Presterice sp. Paracyclogs timbriatus P. fimbri Microcharon (et alignita) Pleco Pleco Pleco Pristine pris	Chaetogaster diaphanus	C.diaph	Spiralix cf vitrea	Spiralix	Niphargus sp. (juveniles)	Niphargus
Nais alpina N.alpi Hydracarina Hydrac Nighargus gr. kochionus N.koch Nais brotota N.barb Aconthocyclaps venustus A.venustus Miphargus srg. kochionus N.Lcaspar Nais brotota N.barb Aconthocyclaps robustus A.venustus Solentinello sp Salentinel Nais groommunis N.com Aconthocyclaps sinitius Solentinello sp Salentinel Nais groommunis N.com Aconthocyclaps sinitius Maibidus Solentinello sp Asellus Nais dir sp Nais sp Paradoclop filmbritus P.fimbri Microcharon of regine Microcharon Styloria lacustris Styla Eucyclaps maruroles E.serrul Parabathynello of stygio Bathynel Pristano aquiseta foreil Pristaeq Diacyclops peticlius D.putealis Leuctra or major Limosoft Tubficinae Tribi Bryocamptus minutus Batylops Ephermeratel usentilos Epherm Haber Bryocamptus minutus Acrassa Baetis sp. Ephermeratelloginata Lignota Lignota Lignota	Chaetogaster diastrophus	C.diast	Pisidium sp.	Pisidium	Nipharaus rhenorhodanensis	N.rheno
Nois brotataN.bartAcanthocyclops renustusAvenustusNahargus capport/N.casparNais bretscheriN.bretAcanthocyclops sensitivusA.sensitivusSolentinello spSalentinellNais grocimunisN.comAcanthocyclops sensitivusA.sensitivusSolentinello spSalentinellNais prodalisN.pardMegocyclops viridisM.viridisProzesflus wolteriP.walteriNois diva spNais pordalisN.pardMegocyclops viridisM.viridisProzesflus wolteriStylaria lacustrisStylaEucyclops serulatusE.serulPricabathynello f stygioBathynelStylaria lacustrisStylaEucyclops serulatusE.serulPricabathynello f stygioBathynelPristina genkinacePristjenkDiacyclops puterolisD.putealisEuctra reginoeLingorTubificinaeTubifBryocamptus viphosB.typhlopsEuctra reginoulatusEigeniculataLageniculataLageniculataLageniculataSignitaLinmodrilus sp.LinmohiAttheyella paraanphus viphosB.typhlopsB.typhlopsLophocheta ignataLignotaAttheyella paraanphus viphosB.typhlopsB.typhlopsPatamathis undivalerisP.moldavPesceus schmellP.schmeHabrelepidies sp.Haber sp.Limochila Attheyella paraanphulaicaA.crassaBaetis sp.Liphocheta ignataLimochilus sp.Limochila Sp.Tubifek Epoctophanes richardiE.schme attheyella paraanphulaikaPelataLiphocheta ig	Nais alpina	N.alpi	Hydracarina	Hydrac	Niphargus gr. kochianus	N.koch
Nois bretscheriN bretAcanthocyclops sobustusArabustusNabrargus griotanusM.fontanNais gri communisN.comAcanthocyclops saludusAsensitivusSalentinello spSalentinelNais gri communisN. ReinMacrocyclops suidusM.atibidusAsellus sp.AsellusNais dar spNais spParacyclops finihatusP.fimbirMicrocharon of regineMicrocharonStylaria lacustrisStylaEucyclops serrulatusE.serrulProzellus walteriPercoharonStylaria lacustrisStylaEucyclops serrulatusE.serrulProzebtra (juveniles)PlecoPristina aequiseta forellPrist, eqDiacyclops belgicusD.belgicusLeuctra majorLinaciaPristina perkineePrist, etc.Diacyclops putcalisD.putcalisLeuctra a fucuralLgericulataHaberRyocomptus minutusB.tminutusEthermeroptare (juveniles)EthereEthereIbindinius p.LinnondriiAttheyella granaphtolicaA.paranHeptagenilae (juveniles)EthereLophochaeta (gnotaLignotaAttheyella granaphtolicaA.paranHeptagenilae (juveniles)P.iuteusPatomotrini veldovskipP.vejdovNitocrella patranaphtolicaA.paranHeptagenilae (juveniles)P.iuteusPatomotrini veldovskipP.vejdovNitocrella patranaphtolicaA.senisSenisSenisLinnodrilus boffmeisteriLinno hodAttheyella granaphtolicaA.senisRabrolepSenisPatomotrini veldovskip	Nais barbata	N.barb	Acanthocyclops venustus	A.venustus	Niphargus casparyi	N.caspar
Nois prosmunisN.comAcenthocyclops sensitivusAsensitivusSalentinello sp.Salentinello sp.Nais protalisN.etinMacrocyclops albidusM.albidusAsellus proAsellusNais portalisN.pardMargacyclops yindisM.viridisProosellus watteriP. watteriNais portalisN.pardMargacyclops yindisM.viridisProsoellus watteriP. watteriNais portalisStylarEucyclops serurulatusE.serrulParabothynello citygiaBattynelVejdovskiella IntermediaVejdovEucyclops parulatusE.macrurPlecopter (juveniles)PlecoPristina jenkineePrist,jenkDiacyclops putealisD.putealisLeuctra of fuscaL fuscaTubificinaeTubifBryocomptus tryphlopsB.typhlopsBattyphlopsBattypisBaetisLinnodrilus sp.LinnodriAttheyella paranaphtalicaA.paranHeptagenidae (juveniles)HeptagenLaphotheta ignataLignotaAttheyella paranaphtalicaA.paranHeptagenidae (juveniles)HeptagenLaphotheta ignataLignotaAttheyella vulneriA.wulimPotamathrik vidovskyiP. tuleusParamorycitae barbatusP. Nitokar hibernicaNitokar hibernicaNitokar hibernicaNitokar hibernicaLaphotheta ignataLignotaAttheyella paranaphtalicaA.paranHeptagenidae (juveniles)HeptagenLaphotheta ignataLignotaAttheyella paranaphtalicaA.paranHeptagenidae (juveniles)HeptagenLaphotheta	Nais bretscheri	N.bret	Acanthocyclops robustus	A.robustus	Niphargus gr fontanus	N.fontan
Nois etinguisN. elinMacrocyclops albidusM.albidusAselius sp.AseliusNais pardalisN. pardMegacyclops viridisM.viridisProacellaw witeriP.walteriNais div spNais.spParacyclops pinbriatusE.serrulParabethynello d'stygiaBathynelStylaria lacustrisStylaEucyclops serrulatusE.serrulParabethynello d'stygiaBathynelPristina aequiseta foreliPrist.aeqDiacyclops belgicusD.belgicusLeutra majorLmajorPristina aequiseta foreliPrist.aeqDiacyclops belgicusD.belgicusLeutra majorLmajorPristina ienkineePrist.lenkDiacyclops puteolisD.putealisLeutra majorLimajorHaber truginiHaber truginiBryocomptus trybhopsB.typhlopsEphemeroptrea (juveniles)EphemLinnodrilus haffmeisteriLinnodriAttheyella paranephtolicaA.crassaBaetis sp.BaetisLinnodrilus haffmeisteriLinno.hofAttheyella paranephtolicaA.paranHeptagenilae (juveniles)HeptagenPotamathrix vidoviskyP.veldovNitocrella paranephtolicaA.paranHeptagenilae (juveniles)P.tetusPotamathrix vidoviskyP.veldovNitocrella paranephtolicaA.paranHeptagenilae (juveniles)P.tetusPatamathrix vidoviskyP.veldovNitocrella paranephtolicaA.paranHeptagenilae (juveniles)P.tetusPatamathrix vidoviskyP.veldovMitorrella paranephtolicaA.paranHeptagenilaeHydrogylicae </td <td>Nais ar communis</td> <td>N.com</td> <td>Acanthocyclops sensitivus</td> <td>A.sensitivus</td> <td>Salentinella sp</td> <td>Salentinel</td>	Nais ar communis	N.com	Acanthocyclops sensitivus	A.sensitivus	Salentinella sp	Salentinel
Nais pardalisN.pardMegacyclops viridisM.viridisProasellus walteriP.walteriNais div spNals.spParacyclops fimbriatusP.fimbriMicrocharon C reginaeMicrocharonStylaria lacustrisStylaEucyclops racrurolesE.serrulPerabathynelia C styglaBattynelVejdovskiella intermeliaVejdovEucyclops racrurolesE.macrurPiecoptera (juveniles)PiecoPristina aequiseta foreriPrist.genDiacyclops belgicusD.belgicusLeuctra majorLmajorPristina finaePrist.jenkDiacyclops belgicusD.putealisLeuctra of fuscaLgeniculataHaber Sp.HaberBryccomptus tryphlopsB.vejdovSerratella gintaSignitaLimnodrilus hoffmeisteriLimnodriAttheyella crassaA.crassaBaetis Sp.BaetisLophochaeta ignotaLignotaAttheyella paranaphtolicaA.paranHeptageniae (juveniles)P. Haber ByPotamathrix woldaviensisP.moldavPesceus schmeiliP.schmeHabroleptoides sp.HabrolepPotamathrix woldaviensisP.moldavPesceus schmeiliNitokra liberaCaenis sp.Caenis sp.Tubles sp.TublesPetrochanes richardiErichardiHydropsy e.cenis sp.CaenisRhyacodrilus sp.RhyacodFontinalicaris fontinalisF.fontinalisCheumatopsyche sp.HydropsyRhyacodrilus sp.RhadoAlona quadrangularisA.quadrangPaycherypia pusillaP.pusillaParastencoSpiridAl	Nais elinauis	N.elin	Macrocyclops albidus	M.albidus	Asellus sp.	Asellus
Nais.spNais.spParacyclops fimbriatusP.fimbriMicrocharon ef reginaeMicrocharonStylaria lacustrisStylaEucyclops seruilatusE.serruiParacyclops iseruilatusE.serruiParacyclops iseruilatusE.serruiParacyclops iseruilatusE.serruiParacyclops iseruilatusE.serruiParacyclops iseruilatusE.serruiParacyclops iseruilatusE.serruiParacyclops iseruilatusE.serruiParacyclops iservicesElevictor anglorLimsica<	Nais pardalis	N.pard	Meaacvclops viridis	M.viridis	Proasellus walteri	P.walteri
Stylaria Eucyclops seruiatus E.serul Parabathynelia Cistygia Bathynel Vejdovskiella intermedia Vejdov Eucyclops seruiatus E.nacrur Plecoptera (juveniles) Pleco Pristina ogenizeta foreil Prista, agenizeta foreil Prista, agenizeta foreil Pleco diversita Leuctra mojor Limajor Tubificinae Prista, agenizeta foreil Diacyclops belejuus D.belgicus Leuctra geniculata Lgeniculata Haber sp. Haber tur Bryocomptus vijdiovskyi B.vejdov Serreta geniculata Lgeniculata Immodrilus sp. Limnodri Attheyella crassa Acrassa Baetis Jumodrilus sp. Beatis Lamnodrilus sp. Limnodri Attheyella vulmeri Avulm Potamathris moldowiensis P.luteus Potamathris moldowiensis P.molda Attheyella vulmeri Avulm Potamathris wejdovskyi P.yeldov Nitokra hibernica Nitokra Caenis sp. P.lavtos Paraten	Nais div sp	Nais sp	Paracyclons fimbriatus	P fimbri	Microcharon cf reginge	Microcharon
Vejdovskiello intermedia Vejdov Eucyclops macruroides E macrur Plecoptera (juveniles) Pleco Pristina aequiseta foreli Pristana (mknae Pristina (mknae Pristina (mknae Pristina (mknae Lusca Leghochacta Legnochacta Legnochacta Pleta Pleta </td <td>Stylaria lacustris</td> <td>Styla</td> <td>Eucyclops serrulatus</td> <td>E.serrul</td> <td>Parabathynella cf styaia</td> <td>Bathynel</td>	Stylaria lacustris	Styla	Eucyclops serrulatus	E.serrul	Parabathynella cf styaia	Bathynel
Pristina aequiseta foreli Prist.aeq Diacyclops belgicus D.belgicus Leuctra major L.major Pristina jenkinae Prist.jenk Diacyclops putealis D.putealis Leuctra of fusca L.fusca Publicinae Tubif Bryocamputs minutus B.minutus Euleuctra geniculata Lgeniculata Haber Bryocamputs windvas B.wipdiov Shemeroptera (juveniles) Ephem Haber turquini Haber Bryocamputs vejdovskyi B.vejdov Serratelia ignita Signita Limnodrilus sp. Limno.hof Attheyella corasa Acrassa Baetis sp. Baetis Lophochaeta ignota Lignota Attheyella paranophtalica A.paran Heptagenidae (juveniles) Heptagen Potamothrix moldoviensis P.moldav Pesceus schmeili P.schme Habrolep Potamothrix luteus P.luteus Potamothrix vejdovskyi P.vejdov Nitokra Iberrica Nitokra Coenis sp. Caenis Tubifex sp. Ramphi Plastano Fontinalis Cheunatopsyche lepida Clepida Polycentropus	Veidovskiella intermedia	Veidov	Eucyclops macruroides	E.macrur	Plecoptera (iuveniles)	Pleco
Pristina jenkinaePrist.jenkDiacyclops putealisD.putealisLeuctra cf fuscaL.fuscaTubificinaeTubifiBryocamptus syninusB.minutusEluetra geniculataL.geniculataHaber sp.Haber turBryocamptus vejdovskyiB.vejdovSerratella ignitaSignitaLimnodriius sp.LimnodriAttheyella crassoA.crassaBaetis sp.BaetisLimnodriius soffLimnodriAttheyella paranaphtilaaA.paranHeptagenilae (juveniles)HeptageniLophochaeta ignotaLignotaAttheyella paranaphtilaA.paranHeptagenilae (juveniles)P.luteusPatamothrix moldaviensisP.noldavPesceus schmeillP.schmeHabrolepHabrolepPatamothrix vejdovskyiP.vejdovNitocrella psammophilaNitocrellaEphemera sp.EphemeraSubifex sp.TubifexEpactophanes richardiE.richardiHydropsyche sp.HydropsyRhyacodrilus sp.RhyacodFontinalicaris fontinalisF.fontinalisCheumatopsyche lepidaC.lepidaRhyacodrilus sanphigenusR.amphiParastenacaris f. glarealaA.gutataHydroptildaeHydropsillaPhalodrilus aphigenusR.falciCladocera (juveniles)CladoPsychomyja pusillaP.gusillaParastenacaris f. glarealaA.gutataHydroptilidaeHydroptilPalladrilinaePhallodrilinaePhalodrilinaeA.gutataHydroptilidaePhalodrilinaePhalodrilinaeA.lona gudanagulariaA.gutataHydroptilidae </td <td>Pristina aequiseta foreli</td> <td>Prist.aeq</td> <td>Diacyclops belgicus</td> <td>D.belgicus</td> <td>Leuctra major</td> <td>L.major</td>	Pristina aequiseta foreli	Prist.aeq	Diacyclops belgicus	D.belgicus	Leuctra major	L.major
TubificinaeTubifBryocamptus minutusB.minutusEuleuctra geniculataL.geniculataHaberBryocamptus typilopsB.typilopsEphemeroptera (juveniles)EphemHaber turquiniHaber turBryocamptus vejdovskyiB.vejdovSerratella ignitaS.ignitaLimnodrilus sp.Limno.hofAttheyella crassaA.crassaBaetis sp.BaetisLimnodrilus hoffmeisteriLimno.hofAttheyella vendovaluA.paranHeptageniidae (juveniles)HeptageniPotamothrix moldaviensisP.moldavPesceus schmeiliP.schmeHabrolephemera spEphemeraPotamothrix wejdovskyiP.vejdovNitocrella psammorphilaNitorcellaEphemera spEphemeraPotamothrix vejdovskyiP.vejdovNitorcella psammorphilaNitorcellaEphemera spCaenisTubifex sp.TubifexEpactophanes richardiE.richardiHydropsyche sp.HydropsyRhyacodrilus sp.RhyacodFontinalicaris fontinalisF.fontinalisCheumatopsyche lepidaClepidaParastenodrilus sp.R.falciCladocera (juveniles)CladoParastenoflavomaculatusPhallodAlona quadrangularisA.quadrangLeptoceridaeLeptoceridPhallodrilinaePhallodAlona ectangulaA.rectangEsolus sp.EsolusPhallodrilinaePhallodAlona ectangulaA.rectangEsolus sp.ElimisEnchytrae idaeEnchytChydorus spheericusC.spigerStenelmis sp.ElimisE	Pristina jenkinae	Prist.jenk	Diacyclops putealis	D.putealis	Leuctra cf fusca	L.fusca
Haber sp.HaberBryocamptus typhlopsB.typhlopsEphemeroptera (juveniles)EphemHaber turquiniHaber.turBryocamptus vejdovskyiB.vejdovSerratella (gnita)S.ignitaLimnodrilus sp.LimnochiAttheyella crassaA.crassaBaetis sp.BaetisLimnodrilus hoffmeisteriLimnochiAttheyella varanaHeptagenildae (juveniles)HeptagenLophochaeta ignotaLignotaAttheyella varanaphtalicaA.paranHeptagenildae (juveniles)HeptagenPotamothrix vejdovskyiP.vejdovNitocrella psammaphilaNitocrellaEphemera sp.EphemeraPotamothrix vejdovskyiP.vejdovNitocrella psammaphilaNitokraCaenis sp.CaenisPamonyctides barbatusP.barbatNitokra hilernicaNitokraCaenis sp.CaenisPhyacodrilus sp.Tubifex sp.Tubifex sp.Parastenocaris of, glareolaParastenoflavomaculatusP.flavomRhyacodrilus amphigenusR.amphiParastenocaris of, glareolaParastenoflavomaculatusP.flavomRhyacodrilus dali falciformisR.falciCladocera (juveniles)CladoPsychomyia pusillaP.pusillaParasthyacodrilus balmensisAlona guatanaA.quadrangLeptoceridaeLeptoceridaePhallodrilinaePhallodAlona veltneriA.weltnLimnius sp.LimniusSpiridion phreaticolaSpiridAlona veltneriA.weltnLimnius sp.LimniusFridericia sp.FrideriPhreaticolaSpiridi <td>Tubificinae</td> <td>Tubif</td> <td>Bryocamptus minutus</td> <td>B.minutus</td> <td>Euleuctra geniculata</td> <td>L.geniculata</td>	Tubificinae	Tubif	Bryocamptus minutus	B.minutus	Euleuctra geniculata	L.geniculata
Haber turquiniHaber.turBryocamptus vejdovskyiB.vejdovSerratella ignitaS. IgnitaLimnodrilus sp.LimnodrinAttheyella crassaA.crassaBætis sp.BætisLaphochaeta ignotaL.ignotaAttheyella paranaphtolicaA.paranHeptagenidae (juveniles)HeptagenLaphochaeta ignotaL.ignotaAttheyella paranaphtolicaA.paranHeptagenidae (juveniles)HeptagenPotamothrix moldaviensisP.moldavPesceus schmelliP.schmeHabroleptoides sp.Habroleptoides sp.Potamothrix vejdovskyiP.vejdovNitokra hibernicaNitokraCaenis sp.CaenisPammoryctides barbatusP.barbatNitokra hibernicaNitokraCaenis sp.CaenisPhyacodrilus sp.TubifexEpactophanes richardiE.richardiHydropsyche sp.HydropsyRhyacodrilus sp.RhyacodFontinalicaris fontinalisF.fontinalisCheumatopsyche lepidaClepidaParasteno drilus ekmaniP.ekmaniAlona guttataAguttataHydroptilludeHydroptilPharantyacdrilus ekmaniP.ekmaniAlona quadrangularisA.guadaragLeptoceridaeLeptoceridPhyacodrilus balmensisNana eetinguuA.rectangEsolus sp.StenelmStenelmSpiridon phreaticolaSpiridAlona rectangulaA.rectangEsolus sp.StenelmCernosvitoviella spCernosvChydorus pigerC.jigerStenelmis sp.StenelmCernosvitoviella sp.FrideriPhreatolona phreatica <td>Haber sp.</td> <td>Haber</td> <td>Bryocamptus typhlops</td> <td>B.typhlops</td> <td>Ephemeroptera (juveniles)</td> <td>Ephem</td>	Haber sp.	Haber	Bryocamptus typhlops	B.typhlops	Ephemeroptera (juveniles)	Ephem
Limnodrillus sp.LimnodriAttheyella crassaA.crassaBaetis sp.BaetisLimnodrilus hoffmeisteriLimno.hofAttheyella paranaphtalicaA.paranHeptagenidae (juveniles)HeptagenLophochaeta ignotaLignotaAttheyella wulmeriA.wulmPotamanthus luteusP. luteusPatamothrix moldaviensisP.moldavPescue schmeiliP.schmeHabroleptoides sp.Habroleptoides sp.Habroleptoides sp.Patamothrix wejdovskyiP.vejdovNitocrella psammophilaNitocrellaEphemera sp.Ephemera sp.EphemeraPsammoryctides barbatusP.barbatNitokra hibernicaNitokraCaenis sp.CaenisHydropsyche sp.Hydropsyche sp.Hydropsyche sp.Hydropsyche sp.Hydropsyche sp.Hydropsyche sp.Hydropsyche sp.F.flavomRhyacodrilus amphigenusR.amphiParostenocaris cf. glareolaParastenoParastenoF.flavomP.flavomRhyacodrilus falciformisR.falciCladocera (juveniles)CladoPsychomyia pusillaP.pusillaParadrhyacodrilus balmensisR.balmensisAlona guadrangularisA.guatranLeptoceridaeLeptoceridaeSpiridion phreaticolaSpiridAlona weltneriA.weltnLimnius sp.LimniusSpiridion phreaticolaSpiridAlona weltneriaCeradop phreaticaPyheradidae div.sp.DitycidFriderica sp.FrideriPhreatalona phreaticaP.phreaticaDytcidae div.sp.DitycidMarionina argenteaMargentOstraccal giuven	Haber turauini	Haber.tur	Brvocamptus veidovskvi	B.veidov	Serratella janita	S.ignita
Limnodrilus hoffmeisteriLimno.hofAttheyella paranaphtalicaA.paranHeptageniidae (juveniles)HeptagenLophochaeta ignotaLignotaAttheyella wulmeriA.wulmPotamonthus luteusP. luteusPotamothrix vejdovskyiP.vejdovNitocrella psammophilaNitocrellaEphemera spEphemera spPatamothrix vejdovskyiP.vejdovNitocrella psammophilaNitokraCaenis sp.CaenisTubifex sp.TubifexEpactophanes richardiErichardiHydropsyche sp.HydropsyRhyacodrilus sp.RhyacodFontinalicaris fontinalisF.fontinalisCheumatopsyche lepidaC.lepidaRhyacodrilus amphigenusR.amphiParastenocaris cf. glareolaParastenoFoltomaP.pusillaPhalodrilus falciformisR.falciCladocera (juveniles)CladoPsychomyia pusillaP.pusillaPhalodrilus balmensisAlona quadrangularisA.quadrangLeptoceridaeLeptoceridaeSpiridio phreaticolaSpiridAlona rectangulaA-rectangEsolus sp.StenelmCernosvitovilla spFrideriPhreatolana phreaticoCyildae div.sp.DitrodicaSpiridion phreaticolaSpiridAlona vertingsitisLicectriCuimnius sp.LimniusEnchytraeidaeEnchytChydorus gplaericusC.sphaerElmis sp.ElmisEnchytraeidaeEnchytChydorus sphaericusC.sphaerElmis sp.DitycidMarionina argentaMargentOstracoda (juveniles)OstracCeratopogo <td>Limnodrilus sp.</td> <td>Limnodri</td> <td>Atthevella crassa</td> <td>A.crassa</td> <td>Baetis sp.</td> <td>Baetis</td>	Limnodrilus sp.	Limnodri	Atthevella crassa	A.crassa	Baetis sp.	Baetis
Lophochaeta ignotaL.ignotaAttheyella wulmeriA.wulmPotamonthus luteusP. luteusPatamothrix moldoviensisP.moldavPesceus schmeiliP.schmeHabroleptoides sp.HabrolepPotamothrix vejdovskyiP.vejdovNitocrella psammophilaNitocrellaEphemera spEphemeraPsammorycitides barbatusP.barbatNitokra hibernicaNitokraCaenis sp.CaenisTubifex sp.TubifexEpactophanes richardiErichardiHydropsyche sp.HydropsyRhyacodrilus sp.RhyacodFontinalicaris fontinalisF.fontinalisCheumatopsyche lepidaC.lepidaPararhyacodrilus amphigenusR.amphiParastenocaris cf. glareolaParastenoflavomaculatusP.flavomRhyacodrilus addifius ekmoniP.ekmaniAlona guutataA.guttataHydroptildaeHydroptilPhallodrilinaePhallodAlona quudrangularisA.guadrangLeptoceridaeLeptoceridSpiridion phreaticolaSpiridAlona rectangulaA.rectangEsolus sp.EsolusSpiridion phreaticolaSpiridPhalonur actirostrisL.rectirOulimnius sp.UlimniusEnchytraeidaeEnchytChydorus pheericaP.phreatiaMiconecta sp.OilimnFridericia sp.Fridericia sp.Fridericia sp.Fidericia sp.StenelmPridericia sp.Friderici Phreatolona phreaticaP.phreatiaOulimnPridericia sp.Fridericia sp.Fidericia sp.OulimnPridericia sp.Fridericia sp	Limnodrilus hoffmeisteri	Limno.hof	Attheyella paranaphtalica	A.paran	Heptageniidae (juveniles)	Heptagen
Potamothrix moldaviensisP.moldavPesceus schmeiliP.schmeHabroleptoides sp.HabrolepPotamothrix vejdovskyiP.vejdovNitocrella psammophilaNitokraCaenis sp.Ephemera spEphemera spPsammoryctides barbatusP.barbatNitokra hibernicaNitokraCaenis sp.Caenis sp.Caenis sp.Tubifex sp.TubifexEpactophanes richardiErichardiHydropsyche sp.HydropsyRhyacodrilus sp.RhyacodFontinalicaris fontinalisF.fontinalisCheumatopsyche lepidaC.lepidaRhyacodrilus amphigenusR.amphiParastenocaris of. glareolaParastenoflavomaculatusP.flavomRhyacodrilus falciformisR.falciCladocera (juveniles)CladoPsychomyia pusillaP.pusillaPhallodrilinaePhallodAlona quadrangularisA.quadrangLeptoceridaeLeptoceridaRhyacodrilus balmensisR.balmensisAlona rectangulaA.rectangEsolus sp.EsolusSpiridion phreaticolaSpiridAlona veltneriA.weltnLimnius sp.UlimniusCernosvitoviella spCernosvChydorus sphaericusC.sphaerElmis sp.StenelmFridericia sp.FrideriPhreatalona phreaticaP.phreaticaDylicidae div. sp.DitycidMarion argenteaMarionPleuroxus sp.Pleurox Micronecta sp.MicronectaEmpidaePropappus volkiPropapPropap Pseudocandona albicansP.albicansEmpidaeEmpidaeHydrodaspiSpirid <t< td=""><td>Lophochaeta ignota</td><td>L.ignota</td><td>Attheyella wulmeri</td><td>A.wulm</td><td>Potamanthus luteus</td><td>P. luteus</td></t<>	Lophochaeta ignota	L.ignota	Attheyella wulmeri	A.wulm	Potamanthus luteus	P. luteus
Potamothrix vejdovskyiP.vejdovNitocrella psammophilaNitocrellaEphemera spEphemeraPsammoryctides barbatusP.barbatNitokra hibernicaNitokraCaenis sp.CaenisTubifex sp.TubifexEpactophanes richardiE.richardiHydropsyche sp.HydropsyRhyacodrilus sp.RhyacodFontinalicaris fontinalisF.fontinalisCheumatopsyche lepidaC.lepidaRhyacodrilus amphigenusR.amphiParastenocaris cf. glareolaParastenoflavomaculatusP.flavomRhyacodrilus falciformisR.falciCladocera (juveniles)CladoPsychomya pusillaP.pusillaPararhyacodrilus ekmaniP.ekmaniAlona guadrangularisA.guttataHydroptilidaeHydroptilPhallodrilinaePhallodAlona quadrangularisA.quadrangLeptoceridaeLeptoceridRhyacodrilus balmensisR.balmensisAlona aveltneriA.weltnLimnius sp.LimniusEnchytraeidaeEnchytChydorus pigerC.pigerStenelmis sp.StenelmCernosvi toviella spCernosvChydorus spheericusC.splaverMicronecta sp.Micronecta sp.Friderica sp.FrideriPhetatlonan arcetitostrisLreetirDylicidae div.sp.DitycidMarionina sp.MarionPleuroxus sp.Pleuroxus sp.Pleuroxus sp.DitycidMarionina sp.MarionPleuroxus sp.Pleuroxus sp.SimulidaeEmpididaeHaplotaxis gordioldesHaplotaxPseudocandona albicansP.albicans<	Potamothrix moldaviensis	P.moldav	Pesceus schmeili	P.schme	Habroleptoides sp.	Habrolep
Psammoryctides barbatusP.barbatNitokra hibernicaNitokraCaenis sp.CaenisTubifex sp.TubifexEpactophanes richardiE.richardiHydropsyche sp.HydropsyRhyacodrilus sp.RhyacodFontinalicaris fontinalisF.fontinalisCheumatopsyche lepidaC.lepidaRhyacodrilus amphigenusR.amphiParastenocaris cf. glareolaParastenoflavomaculatusP.flavomRhyacodrilus falciformisR.falciCladocera (juveniles)CladoPsychomyia pusillaP.pusillaPararhyacodrilus kamaiP.ekmaniAlona guttataA.guttataHydroptillideHydroptillPhallodrilinaePhallodAlona rectangulaA.rectangEsolus sp.EsolusSpiridion phreaticolaSpiridAlona veltneriA.weltnLimnius sp.LimniusEnchytraeidaeEnchytChydorus pigerC.pigerStenelmis sp.StenelmEnchytraeidaeFrideriPhreatolana pheaticaP.pleuroxMicronecta gp.MicronectaMarionina sp.MarigentOstracoda (juveniles)OstracCeratopogCeratopogPropappus volkiPropapPseudocandona albicansP.albicansEmpididaeEmpididaeHydra Sp.HydraMarconada albicansP.albicansEmpididaeEmpididaeIndiversity and the sp.Hydra Sp.Piteratoada (juveniles)OstracCeratopog onidaeCeratopogPropappus volkiPropapPseudocandona albicansP.albicansEmpididaeEmpididae <td>Potamothrix vejdovskyj</td> <td>P.vejdov</td> <td>Nitocrella psammophila</td> <td>Nitocrella</td> <td>Ephemera sp</td> <td>Ephemera</td>	Potamothrix vejdovskyj	P.vejdov	Nitocrella psammophila	Nitocrella	Ephemera sp	Ephemera
Tubifex sp.TubifexEpactophanes richardiE.richardiHydropsyche sp.HydropsyRhyacodrilus sp.RhyacodFontinalicaris fontinalisF.fontinalisClepidaClepidaRhyacodrilus sp.R.amphiParastenocaris cf. glareolaParastenoPolycentropuscfRhyacodrilus falciformisR.falciCladocera (juveniles)CladoPsychomyia pusillaP.flavomParatyacodrilus kemaniP.ekmaniAlona guttataA.guttataHydroptilidaeHydroptiliPhallodrilinaePhallodAlona quadrangularisA.guadrangLeptoceridaeLeptoceridSpiridion phreaticolaSpiridAlona veltneriA.weltnLimnius sp.ElolusSpiridion phreaticolaSpiridAlona veltneriA.weltnLimnius sp.ElmisEnchytraeidaeEnchytChydorus pigerC.spherStenelmis sp.StenelmCernosvitoviella spCernosvChydorus sphaericusC.spherDitycidaMicronecta sp.Friderica sp.FrideriPhreotolona phreaticaP.phreaticaDyticidae div. sp.DitycidMarionina sp.MarionPleuroxus sp.Pleuroxus sp.Pleuroxus sp.EmplidaeEmplidaePropapu svolkiPropapPseudocandona albicansP.albicansEmplidaeEmplidaeHydra sp.HydraMarmocandona zschokkeiM.zcokkeiMicronectaSimulidaeDugesia tigrinaD.tigriSchellencandona riguetraS.triquetraBrachycera predatorsDiptPredat	Psammoryctides barbatus	P.barbat	Nitokra hibernica	Nitokra	Caenis sp.	Caenis
Rhyacodrilus sp.RhyacodFontinalicaris fontinalisF.fontinalisCheumatopsyche lepidaC.lepidaRhyacodrilus sp.R.amphiParastenocaris cf. glareolaParastenoflavomaculatusP.flavomRhyacodrilus falciformisR.falciCladocera (juveniles)CladoPsychomyia pusillaP.pusillaPararhyacodrilus kanziP.ekmaniAlona gutataA.gutataHydroptilidaeHydroptilPhallodrilinaePhallodAlona quadrangularisA.quadrangLeptoceridaeLeptoceridRhyacodrilus balmensisR.balmensisAlona rectangulaA.rectangEsolus sp.EsolusSpiridion phreaticolaSpiridAlona weltneriA.weltnLimnius sp.LimniusEnchytraeidaeEnchytChydorus phreaticusC.spiaerC.pigerStenelmis sp.StenelmFridericia sp.FrideriPhreatalona phreaticaP.phreaticaDyticide div.sp.DitycidMarionina sp.MarionPleuroxus sp.PleuroxMicronecta sp.MicronectaMarionina argenteaM.argentOstracoda (juveniles)OstracCeratopogonidaeCeratopogPropapus volkiPropapPseudocandona albicansP.albicansEmplitidaeTipulidaeHydraMarionana zschokkeiM.zschokSimulidaeSimulidaeDugesia agoncephalaD.gonoCryptocandona kieferiC.kieferiTipulidaeDugesia tigrinaD.tigriSchellencandona triquetraStriquetraStriquetraPhagocWegelini	Tubifex sp.	Tubifex	Epactophanes richardi	E.richardi	Hydropsyche sp.	Hydropsy
DescriptionDescriptionPolycentropusCfRhyacodrilus amphigenusR.amphiParastenocoris cf. glareolaParastenoflavomaculatusP.flavomRhyacodrilus falciformisR.falciCladocera (juveniles)CladoPsychomyia pusillaP.pusillaPararhyacodrilus falciformisR.falciCladocera (juveniles)CladoPsychomyia pusillaP.pusillaPhallodrilinaePhallodAlona guatarangularisA.guataaHydroptilidaeHydroptilPhallodrilinaePhallodAlona quatarangularisA.guatarangLeptoceridaeLeptoceridRhyacodrilus balmensisR.balmensisAlona rectangulaA.rectangEsolus sp.EsolusSpiridion phreaticolaSpiridAlona weltneriA.weltnLimnius sp.LimniusEnchytraeus buchholziE.buchLathonura rectirostrisL.rectirOulimnius sp.OulimnFridericia sp.FrideriPhreatalona phreaticaP.phreaticaDyticidae div. sp.DitycidMarionina sp.MarionPleuroxus sp.PleuroxMicronecta sp.MicronectaPropappus volkiPropapPseudocandona albicansP.albicansEmpidiaeEmpidiaeHaplotaxPseudocandona aschokkeiM.zschokSimulidaeSimulidaeDugesia gonocephalaD.gonoCryptocandona kieferiC.kieferiTipulidaeDugesia tigrinaD.tigriSchellencandona figuetraS.triquetraBrachycera predatorsPhagocata sp.PhagocVergeliniF.wegel	Rhvacodrilus sp.	Rhvacod	Fontinalicaris fontinalis	F.fontinalis	Cheumatopsyche lepida	C.lepida
Rhyacodrilus amphigenusR.amphiParastenocaris cf. glareolaParastenoflavomaculatusP.flavomRhyacodrilus falciformisR.falciCladocera (juveniles)CladoPsychomyia pusillaP.pusillaParantyacodrilus ekmaniP.ekmaniAlona guttataA.guttataHydroptilladeHydroptilPhallodrilinaePhallodAlona guadrangularisA.guadrangLeptoceridaeLeptoceridaeRhyacodrilus balmensisR.balmensisAlona quadrangularisA.quadrangLeptoceridaeLeptoceridaeSpiridion phreaticolaSpiridAlona weltneriA.weltnLimnius sp.LimniusEnchytraeidaeEnchytChydorus pigerC.sphaerC.pigerStenelmis sp.StenelmCernosvitoviella spCernosvChydorus sphaericusC.sphaerDulimnius sp.OulimnFriderica sp.FrideriPhreatalona phreaticaP.phreaticaDyticidae div. sp.DitycidMarionina argenteaM.argentOstracoda (juveniles)OstracCeratopogonidaeCeratopogPropapus volkiPropapPseudocandona archicaP.amarchicaLimonidaeLimonidaeHydra sp.HydraMarmocandona zschokkeiM.zschokSimulidaeSimulidaeDugesia gonocephalaD.gonoCryptocandona kieferiC.kieferiTipulidaeTipulidaeDugesia tigrinaD.tigriSchellencandona triguetraS.triquetraBrachycera predatorsDiptPredatPhagocata sp.PhagocWergeliniF.wegeliniF.w		,	,		Polvcentropus cf	
Rhyacodrilus falciformisR.falciCladocera (juveniles)CladoPsychomyia pusillaP.pusillaParantyacodrilus ekmaniP.ekmaniAlona guttataA.guttataHydroptilidaeHydroptilPhallodrilinaePhallodAlona quadrangularisA.guttataHydroptilidaeHydroptilRhyacodrilus balmensisR.balmensisAlona rectangulaA.rectangEsolus sp.EsolusSpiridion phreaticolaSpiridAlona weltneriA.weltnLimnius sp.LimniusEnchytraeidaeEnchytChydorus pigerC.pigerStenelmis sp.StenelmCernosvitoviella spCernosvChydorus sphaericusC.sphaerElmis sp.OulimnFridericia sp.FrideriPhreatalona phreaticaP.phreaticaDyticidae div. sp.OulimnFridericia sp.KridenOstracoda (juveniles)OstracCeratopogonidaeCeratopogMarionina argenteaM.argentOstracoda (juveniles)OstracCeratopogonidaeEmpididaeHaplotaxis gordioidesHaplotaxPseudocandona antrchicaP.amarchicaLimoniidaeLimoniidHydra sp.HydraMarconandona zschokkeiM.zschokSimulidaeSimulidaeSimulidaeDugesia gionacephalaD.gonoCryptocandona kieferiC.kieferiTipulidaeNymChirPhagocata sp.PhagocWergotona kieferiC.kieferiNymphe of ChironomidaeNymChirPhagocata sp.PhagoPenenis candonaCrandidaCcandidaOrthocladiinae	Rhyacodrilus amphigenus	R.amphi	Parastenocaris cf. glareola	Parasteno	flavomaculatus	P.flavom
Pararhyacodrilus ekmaniP.ekmaniAlona guttataA.guttataHydroptilidaeHydroptilPhallodrilinaePhallodAlona quadrangularisA.guadrangLeptoceridaeLeptoceridaRhyacodrilus balmensisR.balmensisAlona rectangulaA.rectangEsolus sp.EsolusSpiridion phreaticolaSpiridAlona weltneriA.weltnLimnius sp.LimniusEnchytraeidaeEnchytChydorus pigerC.pigerStenelmis sp.StenelmCernosvitoviella spCernosvChydorus sphaericusC.sphaerElmis sp.OulimnFridericia sp.FrideriPhreatolona phreaticaP.phreaticaDyticidae div. sp.DitycidMarionina sp.MarionPleuroxus sp.PleuroxMicronecta sp.MicronectaPropapus volkiPropapPseudocandona albicansP.albicansEmpidiaeEmpidiaeHydra sp.HydraMarmocandona zschokkeiM.zschokSimulidaeSimulidaeDugesia gonocephalaD.gonoCryptocandona kieferiC.kieferiTipulidaeNigrodiaePolycelis nigraP.nigraCandona candidaC.candidaOrthocladinaeOrthocladiPolycelis tenuisP.tenuisCandona candidaC.candidaOrthocladinaeOrthocladMarionina sp.BragotaCryptocandona kieferiC.kieferiTipulidaeTipulidaeHydra sp.HydraMarmocandona zschokkeiM.zschokSimulidaeSimulidaeDugesia tigrinaD.tigriSchellencandona	Rhyacodrilus falciformis	R.falci	Cladocera (juveniles)	Clado	Psychomyia pusilla	P.pusilla
PhallodrilinaePhallodAlona quadrangularisA.quadrangLeptoceridaeLeptoceridaRhyacodrilus balmensisR.balmensisAlona rectangulaA.rectangEsolus sp.EsolusSpiridion phreaticolaSpiridAlona weltneriA.weltnLimnius sp.LimniusEnchytraeidaeEnchytChydorus pigerC.pigerStenelmis sp.StenelmCernosvitoviella spCernosvChydorus parericusC.sphaerElmis sp.ElmisEnchytraeus buchholziE.buchLathonura rectirostrisL.rectirOulimnius sp.OulimnFridericia sp.FrideriPhreatalona phreaticaP.phreaticaDyticidae div. sp.DitycidMarionina sp.MarionPleuroxus sp.PleuroxMicronecta sp.MicronectaMarionina argenteaM.argentOstracoda (juveniles)OstracCeratopogonidaeCeratopogPropappus volkiPropapPseudocandona albicansP.albicansEmpididaeEmpididaeHydra sp.HydraMarmocandona zschokkeiM.zschokSimulidaeSimulidaeDugesia gonocephalaD.gonoCryptocandona kieferiC.sirdiuetraBrackycera predatorsDiptPredatPhagocat sp.PhagocwegeliniF.wegeliniNymphe of ChironomidaeNymChirPolycelis tenuisP.nigraCandona anglectaC.candidaOrthocladiinaeOrthocladiDugesia tigrinaP.ingraCandona neglectaC.canglectaCorinoneura sp.CorinoneuraPhagocat sp.<	Pararhyacodrilus ekmani	P.ekmani	Alona guttata	A.guttata	Hydroptilidae	Hydroptil
Rhyacodrilus balmensisR.balmensisAlona rectangulaA.rectangEsolus sp.EsolusSpiridion phreaticolaSpiridAlona weltneriA.weltnLimnius sp.LimniusEnchytraeidaeEnchytChydorus pigerC.pigerStenelmis sp.StenelmCernosvitoviella spCernosvChydorus sphaericusC.sphaerElmis sp.ElmisEnchytraeus buchholziE.buchLathonura rectirostrisLrectirOulimnius sp.OulimnFridericia sp.FrideriPhreatalona phreaticaP.phreaticaDyticidae div. sp.DitycidMarionina sp.MarionPleuroxus sp.PleuroxMicronecta sp.MicronectaMarionina argenteaM.argentOstracoda (juveniles)OstracCeratopogonidaeCeratopogPropappus volkiPropapPseudocandona albicansP.albicansEmpididaeEmpididaeHydra sp.HydraMarmocandona zschokkeiM.zschokSimulidaeSimulidaeDugesia gonocephalaD.gonoCryptocandona kieferiC.kieferiTipulidaeTipulidaeDugesia tigrinaD.tigriSchellencandona triquetraS.triquetraBrachycera predatorsDiptPredatPhagocata sp.PhagocwegeliniF.wegeliniNymphe of ChironomidaeNymChirPolycelis tenuisP.nigraCandona anglectaC.neglectaCorinoneura sp.CorinoneuraPolycelis tenuisP.tenuisCandona anglectaC.ophthTanypodinaeTanypadinaeErpobdella cf octocul	Phallodrilinae	Phallod	Alona quadrangularis	A.quadrang	Leptoceridae	Leptocerid
Spiridion phreaticolaSpiridAlona weltneriA.weltnLimnius sp.LimniusEnchytraeidaeEnchytChydorus pigerC.pigerStenelmis sp.StenelmCernosvitoviella spCernosvChydorus sphaericusC.sphaerElmis sp.ElmisEnchytraeus buchholziE.buchLathonura rectirostrisL.rectirOulimnius sp.OulimnFridericia sp.FrideriPhreatalona phreaticaP.phreaticaDyticidae div. sp.DitycidMarionina sp.MarionPleuroxus sp.PleuroxMicronecta sp.MicronectaMarionina argenteaM.argentOstracoda (juveniles)OstracCeratopogonidaeCeratopogPropappus volkiPropapPseudocandona albicansP.albicansEmpididaeEmpididaeHaydra sp.HydraMarmocandona zschokkeiM.zschokSimulidaeLimoniidDugesia gonocephalaD.gonoCryptocandona kieferiC.kieferiTipulidaeTipulidaeDugesia tigrinaD.tigriSchellencandona triquetraS.triquetraBrachycera predatorsDiptPredatPhagocata sp.PhagocwegeliniF.wegeliniNymphe of ChironomidaeNymChirPolycelis tenuisP.nigraCandona candidaC.candidaOrthocladinaeOrthocladPolycelis tenuisP.tenuisCandona naglectaC.neglectaCorinoneura sp.Corinoneura sp.Polycelis tenuisP.tenuisCandona naglectaC.ophthTanypodinaeTanypodinaeFrabaeformiscandona<	Rhyacodrilus balmensis	R.balmensis	Alona rectangula	A.rectang	Esolus sp.	Esolus
EnchytraeidaeEnchytChydorus pigerC.pigerStenelmis sp.StenelmiCernosvitoviella spCernosvChydorus sphaericusC.sphaerElmis sp.ElmisEnchytraeus buchholziE.buchLathonura rectirostrisL.rectirOulimnius sp.OulimnFridericia sp.FrideriPhreatalona phreaticaP.phreaticaDyticidae div. sp.DitycidMarionina sp.MarionPleuroxus sp.PleuroxMicronecta sp.MicronectaMarionina argenteaM.argentOstracoda (juveniles)OstracCeratopogonidaeCeratopogPropappus volkiPropapPseudocandona albicansP.albicansEmpididaeEmpididaeHaplotaxis gordioidesHaplotaxPseudocandona zschokkeiM.zschokSimulidaeSimulidaeDugesia gonocephalaD.gonoCryptocandona kieferiC.kieferiTipulidaeTipulidaePhagocata sp.PhagowegeliniF.wegeliniNymphe of ChironomidaeNymChirPolycelis nigraP.nigraCandona candidaC.candidaOrthocladiinaeOrthocladiPolycelis nigraP.tenuisCandona neglectaC.neglectaCorinoneura sp.CorinoneuraNematodaNematoCypria ophthalmicaC.ophthTanytarsiniTanytarsiniGlossiphonia sp.GlossiphPrionocypris zenkeriP.zenkeriChironominiChironomini	Spiridion phreaticola	Spirid	Alona weltneri	A.weltn	Limnius sp.	Limnius
Cernosvitoviella spCernosvChydorus sphaericusC.sphaerElmis sp.ElmisEnchytraeus buchholziE.buchLathonura rectirostrisL.rectirOulimnius sp.OulimniFridericia sp.FrideriPhreatalona phreaticaP.phreaticaDyticidae div. sp.DitycidMarionina sp.MarionPleuroxus sp.PleuroxMicronecta sp.MicronectaMarionina argenteaM.argentOstracoda (juveniles)OstracCeratopogonidaeCeratopogPropappus volkiPropapPseudocandona albicansP.albicansEmpididaeEmpididaeHaplotaxis gordioidesHaplotaxPseudocandona marchicaP.marchicaLimoniidaeSimulidaeDugesia gonocephalaD.gonoCryptocandona kieferiC.kieferiTipulidaeTipulidaeDugesia tigrinaD.tigriSchellencandonaF.wegeliniNymphe of ChironomidaeNymChirPolycelis nigraP.nigraCandona candidaC.candidaOrthocladinaeOrthocladinaePolycelis nigraP.tenuisCandona neglectaC.ophthTanypodinaeTanypodinaeErpobdella cf octoculataErpobdIlyocypris bradyiI.bradyiTanytarsiniTanytarsiniGlossiphonia sp.GlossiphPrionocypris zenkeriP.zenkeriChironominiChironomini	Enchytraeidae	Enchyt	Chydorus piger	C.piger	Stenelmis sp.	Stenelm
Enchytraeus buchholziE.buchLathonura rectirostrisL.rectirOulimnius sp.OulimniusFridericia sp.FrideriPhreatalona phreaticaP.phreaticaDyticidae div. sp.DitycidMarionina sp.MarionPleuroxus sp.PleuroxMicronecta sp.MicronectaMarionina argenteaM.argentOstracoda (juveniles)OstracCeratopogonidaeCeratopogPropappus volkiPropapPseudocandona albicansP.albicansEmpididaeEmpididaeHaplotaxis gordioidesHaplotaxPseudocandona zschokkeiM.zschokSimulidaeSimulidaeJugesia gonocephalaD.gonoCryptocandona kieferiC.kieferiTipulidaeTipulidaeDugesia tigrinaD.tigriSchellencandona candidaS.triquetraBrachycera predatorsDiptPredatPolycelis nigraP.nigraCandona candidaC.candidaOrthocladiinaeOrthocladiPolycelis tenuisP.tenuisCandona neglectaC.neglectaCorinoneura sp.CorinoneuraNematodaNematoLypia ophthalmicaC.ophthTanypodinaeTanypodinaeErpobdella cf octoculataErpobdIlyocypris zenkeriP.zenkeriP.zenkeriChironominiChironomini	Cernosvitoviella sp	Cernosv	Chydorus sphaericus	C.sphaer	<i>Elmis</i> sp.	Elmis
Fridericia sp.FrideriPhreatalona phreaticaP.phreaticaDyticidae div. sp.DitycidMarionina sp.MarionPleuroxus sp.PleuroxMicronecta sp.MicronectaMarionina argenteaM.argentOstracoda (juveniles)OstracCeratopogonidaeCeratopogPropappus volkiPropapPseudocandona albicansP.albicansEmpididaeEmpididaeHaplotaxis gordioidesHaplotaxPseudocandona marchicaP.marchicaLimoniidaeLimoniidHydra sp.HydraMarmocandona zschokkeiM.zschokSimulidaeSimulidaeSimulidaeDugesia gonocephalaD.gonoCryptocandona kieferiC.kieferiTipulidaeTipulidaeDugesia tigrinaD.tigriSchellencandona triquetraS.triquetraBrachycera predatorsDiptPredatPhagocata sp.PhagocP.nigraCandona candidaC.candidaOrthocladiinaeOrthocladPolycelis ingraP.nigraCandona neglectaC.neglectaCorinoneura sp.CorinoneuraNematodaNematoCypria ophthalmicaC.ophthTanypodinaeTanypodinaeErpobdella cf octoculataErpobdIlyocypris bradyiI.bradyiTanytarsiniTanytarsiniGlossiphonia sp.GlossiphPrionocypris zenkeriP.zenkeriChironominiChironomini	Enchytraeus buchholzi	E.buch	Lathonura rectirostris	L.rectir	<i>Oulimnius</i> sp.	Oulimn
Marionina sp.MarionPleuroxus sp.PleuroxMicronecta sp.MicronectaMarionina argenteaM.argentOstracoda (juveniles)OstracCeratopogonidaeCeratopogPropappus volkiPropapPseudocandona albicansP.albicansEmpididaeEmpididaeHaplotaxis gordioidesHaplotaxPseudocandona marchicaP.marchicaLimoniidaeLimoniidHydra sp.HydraMarmocandona zschokkeiM.zschokSimulidaeSimulidaeDugesia gonocephalaD.gonoCryptocandona kieferiC.kieferiTipulidaeTipulidaeDugesia tigrinaD.tigriSchellencandona triquetraS.triquetraBrachycera predatorsDiptPredatPhagocata sp.PhagocPhagocCandona candidaC.candidaOrthocladiinaeOrthocladPolycelis nigraP.nigraCandona neglectaC.neglectaCorinoneura sp.CorinoneuraNematodaNematoCypria ophthalmicaC.ophthTanypodinaeTanypodinaeErpobdella cf octoculataErpobdIlyocypris bradyiI.bradyiTanytarsiniTanytarsini	Fridericia sp.	Frideri	Phreatalona phreatica	P.phreatica	Dyticidae div. sp.	Ditycid
Marionina argenteaM.argentOstracoda (juveniles)OstracCeratopogonidaeCeratopogPropappus volkiPropapPseudocandona albicansP.albicansEmpididaeEmpididaeHaplotaxis gordioidesHaplotaxPseudocandona marchicaP.marchicaLimoniidaeLimoniidHydra sp.HydraMarmocandona zschokkeiM.zschokSimulidaeSimulidaeDugesia gonocephalaD.gonoCryptocandona kieferiC.kieferiTipulidaeTipulidaeDugesia tigrinaD.tigriSchellencandona triquetraS.triquetraBrachycera predatorsDiptPredatPhagocata sp.PhagocPhagocExadona anglectaC.candidaOrthocladiinaeOrthocladPolycelis nigraP.nigraCandona neglectaC.neglectaCorinoneura sp.CorinoneuraNematodaNematoCypria ophthalmicaC.ophthTanypodinaeTanypodinaeErpobdella cf octoculataErpobdIlyocypris bradyiI.bradyiTanytarsiniTanytarsiniGlossiph onia sp.GlossiphPrionocypris zenkeriP.zenkeriChironominiChironomini	Marionina sp.	Marion	Pleuroxus sp.	Pleurox	Micronecta sp.	Micronecta
Propappus volkiPropapPseudocandona albicansP.albicansEmpididaeEmpididaeHaplotaxis gordioidesHaplotaxPseudocandona marchicaP.marchicaLimoniidaeLimoniidaHydra sp.HydraMarmocandona zschokkeiM.zschokSimulidaeSimulidaeDugesia gonocephalaD.gonoCryptocandona kieferiC.kieferiTipulidaeTipulidaeDugesia tigrinaD.tigriSchellencandona triquetraS.triquetraBrachycera predatorsDiptPredatPhagocata sp.PhagocFabaeformiscandonaF.wegeliniNymphe of ChironomidaeNymChirPolycelis nigraP.nigraCandona candidaC.candidaOrthocladiinaeOrthocladPolycelis tenuisP.tenuisCandona neglectaC.ophthTanypodinaeTanypodinaeErpobdella cf octoculataErpobdIlyocypris bradyiI.bradyiTanytarsiniTanytarsiniGlossiphonia sp.GlossiphPrionocypris zenkeriP.zenkeriChironominiChironomini	Marionina argentea	M.argent	Ostracoda (juveniles)	Ostrac	Ceratopogonidae	Ceratopog
Haplotaxis gordioidesHaplotaxPseudocandona marchicaP.marchicaLimoniidaeLimoniidaeHydra sp.HydraMarmocandona zschokkeiM.zschokSimulidaeSimulidaeDugesia gonocephalaD.gonoCryptocandona kieferiC.kieferiTipulidaeTipulidaeDugesia tigrinaD.tigriSchellencandona triquetraS.triquetraBrachycera predatorsDiptPredatPhagocata sp.PhagocFabaeformiscandona wegeliniF.wegeliniNymphe of ChironomidaeNymChirPolycelis nigraP.nigraCandona candidaC.candidaOrthocladiinaeOrthocladPolycelis tenuisP.tenuisCandona neglectaC.neglectaCorinoneura sp.CorinoneuraNematodaNematoCypria ophthalmicaC.ophthTanypodinaeTanypodinaeErpobdella cf octoculataErpobdIlyocypris bradyiI.bradyiTanytarsiniTanytarsiniGlossiph onia sp.GlossiphPrionocypris zenkeriP.zenkeriChironominiChironomini	Propappus volki	Propap	Pseudocandona albicans	P.albicans	Empididae	Empididae
Hydra sp.HydraMarmocandona zschokkeiM.zschokSimulidaeSimulidaeDugesia gonocephalaD.gonoCryptocandona kieferiC.kieferiTipulidaeTipulidaeDugesia tigrinaD.tigriSchellencandona triquetraS.triquetraBrachycera predatorsDiptPredatPhagocata sp.PhagocFabaeformiscandona wegeliniF.wegeliniNymphe of ChironomidaeNymChirPolycelis nigraP.nigraCandona candidaC.candidaOrthocladiinaeOrthocladPolycelis tenuisP.tenuisCandona neglectaC.neglectaCorinoneura sp.CorinoneuraNematodaNematoCypria ophthalmicaC.ophthTanypodinaeTanypodinaeErpobdella cf octoculataErpobdIlyocypris bradyiI.bradyiTanytarsiniTanytarsiniGlossiphonia sp.GlossiphPrionocypris zenkeriP.zenkeriChironominiChironomini	Haplotaxis gordioides	Haplotax	Pseudocandona marchica	P.marchica	Limoniidae	Limoniid
Dugesia gonocephalaD.gonoCryptocandona kieferiC.kieferiTipulidaeTipulidaeDugesia tigrinaD.tigriSchellencandona triquetraS.triquetraBrachycera predatorsDiptPredatPhagocata sp.PhagocFabaeformiscandona wegeliniF.wegeliniNymphe of ChironomidaeNymChirPolycelis nigraP.nigraCandona candidaC.candidaOrthocladiinaeOrthocladPolycelis tenuisP.tenuisCandona neglectaC.neglectaCorinoneura sp.CorinoneuraNematodaNematoCypria ophthalmicaC.ophthTanypodinaeTanypodinaeErpobdella cf octoculataErpobdIlyocypris bradyiI.bradyiTanytarsiniTanytarsiniGlossiphonia sp.GlossiphPrionocypris zenkeriP.zenkeriChironominiChironomini	Hydra sp.	Hydra	Marmocandona zschokkei	M.zschok	Simulidae	Simulidae
Dugesia tigrinaD.tigriSchellencandona triquetraS.triquetraBrachycera predatorsDiptPredatPhagocata sp.PhagocFabaeformiscandona wegeliniF.wegeliniNymphe of ChironomidaeNymChirPolycelis nigraP.nigraCandona candidaC.candidaOrthocladiinaeOrthocladPolycelis tenuisP.tenuisCandona neglectaC.neglectaCorinoneura sp.CorinoneuraNematodaNematoCypria ophthalmicaC.ophthTanypodinaeTanypodinaeErpobdella cf octoculataErpobdIlyocypris bradyiI.bradyiTanytarsiniTanytarsiniGlossiphonia sp.GlossiphPrionocypris zenkeriP.zenkeriChironominiChironomini	Dugesia gonocephala	D.gono	Cryptocandona kieferi	C.kieferi	Tipulidae	Tipulidae
Phagocata sp.Fabaeformiscandona wegeliniF.wegeliniNymphe of ChironomidaeNymChirPolycelis nigraP.nigraCandona candidaC.candidaOrthocladiinaeOrthocladPolycelis tenuisP.tenuisCandona neglectaC.neglectaCorinoneura sp.CorinoneuraNematodaNematoCypria ophthalmicaC.ophthTanypodinaeTanypodinaeErpobdella cf octoculataErpobdIlyocypris bradyiI.bradyiTanytarsiniTanytarsiniGlossiphonia sp.GlossiphPrionocypris zenkeriP.zenkeriChironominiChironomini	Dugesia tigrina	D.tigri	Schellencandona triquetra	S.triquetra	Brachycera predators	DiptPredat
Phagocata sp.PhagocwegeliniF.wegeliniNymphe of ChironomidaeNymChirPolycelis nigraP.nigraCandona candidaC.candidaOrthocladiinaeOrthocladPolycelis tenuisP.tenuisCandona neglectaC.neglectaCorinoneura sp.CorinoneuraNematodaNematoCypria ophthalmicaC.ophthTanypodinaeTanypodinaeErpobdella cf octoculataErpobdIlyocypris bradyiI.bradyiTanytarsiniTanytarsiniGlossiphonia sp.GlossiphPrionocypris zenkeriP.zenkeriChironominiChironomini			Fabaeformiscandona			
Polycelis nigraP.nigraCandona candidaC.candidaOrthocladiinaeOrthocladPolycelis tenuisP.tenuisCandona neglectaC.neglectaCorinoneura sp.CorinoneuraNematodaNematoCypria ophthalmicaC.ophthTanypodinaeTanypodinaeErpobdella cf octoculataErpobdIlyocypris bradyiI.bradyiTanytarsiniTanytarsiniGlossiphonia sp.GlossiphPrionocypris zenkeriP.zenkeriChironominiChironomini	Phagocata sp.	Phagoc	wegelini	F.wegelini	Nymphe of Chironomidae	NymChir
Polycelis tenuisP.tenuisCandona neglectaC.neglectaCorinoneura sp.CorinoneuraNematodaNematoCypria ophthalmicaC.ophthTanypodinaeTanypodinaeErpobdella cf octoculataErpobdIlyocypris bradyiI.bradyiTanytarsiniTanytarsiniGlossiphonia sp.GlossiphPrionocypris zenkeriP.zenkeriChironominiChironomini	Polycelis nigra	P.nigra	Candona candida	C.candida	Orthocladiinae	Orthoclad
NematodaNematoCypria ophthalmicaC.ophthTanypodinaeTanypodinaeErpobdella cf octoculataErpobdIlyocypris bradyiI.bradyiTanytarsiniTanytarsiniGlossiphonia sp.GlossiphPrionocypris zenkeriP.zenkeriChironominiChironomini	Polycelis tenuis	P.tenuis	Candona neglecta	C.neglecta	Corinoneura sp.	Corinoneura
Erpobdella cf octoculataErpobdIlyocypris bradyiI.bradyiTanytarsiniTanytarsiniGlossiphonia sp.GlossiphPrionocypris zenkeriP.zenkeriChironominiChironomini	Nematoda	Nemato	Cypria ophthalmica	C.ophth	Tanypodinae	Tanypodinae
Glossiphonia sp. Glossiph Prionocypris zenkeri P.zenkeri Chironomini Chironomini	Erpobdella cf octoculata	Freehd	Ilvocupris bradui	I bradvi	Tanytarsini	Tanytarsini
		Егрора	nyocypris bruuyi	iuuyi	ranycaronn	i any caronin

1368

- 1370 Table S2. Results of two-ways ANOVA between depths (-20, -40 and -60cm) and hydrological patterns
- 1371 (downwelling, upwelling, no exchange) for abundances and richness of the total hyporheic fauna, the
- 1372 temporary and the permanent hyporheos and the stygobites, after log(x+1) transformation.

1373 Total richness of the hyporheos

Effects	DF	F	р
Depths	2	15.61	0.000002
Hydrological patterns	2	12.96	0.000013
Hydro x depths	4	1.11	n.s.

1374

1375 Total abundances of the hyporheos

Effects	DF	F	р
Depths	2	20.88	< 10 ⁻⁵
Hydrological patterns	2	15.72	0.000002
Hydro x depths	4	0.79	n.s.

1376

1377 Richness of the temporary hyporheos

Effects	DF	F	р
Depths	2	19.17	< 10 ⁻⁵
Hydrological patterns	2	20.19	< 10 ⁻⁵
Hydro x depths	4	0.38	n.s.

1378

1379 Abundances of the temporary hyporheos

Effects	DF	F	р
Depths	2	23.17	< 10 ⁻⁵
Hydrological patterns	2	30.63	< 10 ⁻⁵
Hydro x depths	4	0.61	n.s.

1380

1381 Richness of the permanent hyporheos

Effects	DF	F	р
Depths	2	18.65	< 10 ⁻⁵
Hydrological patterns	2	8.61	0.0004
Hydro x depths	4	0.65	n.s.

1382

1383 Abundances of the permanent hyporheos

		-	
Effects	DF	F	р
Depths	2	15.41	0.000002
Hydrological patterns	2	7.26	0.001
Hydro x depths	4	0.34	n.s.

1384

1385 Richness of the stygobites

70			
Effects	DF	F	р
Depths	2	0.028	n.s.
Hydrological patterns	2	16.34	0.000001
Hydro x depths	4	0.52	n.s.

1386

1387 Abundances of the stygobites

Effects	DF	F	р
Depths	2	0.056	n.s.

Hydrological patterns	2	6.64	0.002
Hydro x depths	4	0.38	n.s.

1389 Abundances of the sediment feeders

Effects	DF	F	р
Depths	2	18.03	< 10 ⁻⁵
Hydrological patterns	2	15.79	0.000002
Hydro x depths	4	0.94	n.s.

1391 Abundances of the POM feeders

Effects	DF	F	р
Depths	2	23.45	< 10 ⁻⁵
Hydrological patterns	2	11.07	0.00005
Hydro x depths	4	0.31	n.s.

1393 Abundances of the Algae feeders

Effects	DF	F	р
Depths	2	19.31	< 10 ⁻⁵
Hydrological patterns	2	13.04	0.000012
Hydro x depths	4	0.70	n.s.

1395 Abundances of the predators

Effects	DF	F	р
Depths	2	11.34	0.00004
Hydrological patterns	2	9.05	0.0003
Hydro x depths	4	0.11	n.s.

1397 Ecological groups' diversity

Effects	DF	F	р
Depths	2	0.42	n.s.
Hydrological patterns	2	8.11	0.0006
Hydro x depths	4	0.19	n.s.

1399 Food-type groups' diversity

Effects	DF	F	р
Depths	2	5.25	0.007
Hydrological patterns	2	4.15	0.019
Hydro x depths	4	1.23	n.s.

1406 Supplementary material: Figures

Fig. S1 : Biplots for hydrological patterns.



- 1412 Fig. S2 : Longitudinal profiles of temperature, electric conductivity, dissolved oxygen, vertical hydraulic
- 1413 gradient and POM (mean values per stations, all depths and all replicated cumulated)



- 1415 Fig. S3a: Frequences of temporary hyporheic taxa according to local hydrology



1421 Fig. S3b: Frequences of permanent hyporheic taxa according to local hydrology







1427Fig. S4: Effects of sediment grain size (left column), sediment stability (central column) and hydrology1428(right column) on taxonomic richness, abundances and functional group abundances of the hyporheic

assemblages. Results of one-way ANOVAs on classes after log-transformation, using stations asreplicates. Mean values and error-types.



Fig. S5: Correlations between abundances and taxonomic richness and local characteristics (Temperature and Dissolved Oxygen) at 60cm depth in downwelling zones.





When considering the relationship of the abundances (upper pannels) and taxonomic richness (lower pannels) with temperature and Oxygen at -60 cm depth in downwelling zones, we observed similar patterns with increasing number of species and individuals with increasing temperature (with maximum values above 25°C) and with available Oxygen (with maximum values above 7 mg.L⁻¹). The hyporheic fauna developp abundant and diversifies assemblages in strong downwelling zones (with warm and well oxygenated water).